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Reviewing the reported pharmacognostic and pharmacological investigations on *Tecoma stans* Juss. ex Kunth

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ARTICLEINFO	A B S T R A C T		
Article Type: Review	This review is destined for a comprehensive assessment of the phytochemistry and medicinal properties of <i>Tecoma stans</i> , a widely used plant in folk cultures, as a traditionally safe and		
<i>Article History:</i> Received: 7 August 2022 Accepted: 14 October 2022	effective treatment for different diseases and complications. The attainable and reachable sources of <i>T. stans</i> confirmed its origin, ethnopharmacological properties, and therapeuti medicinal uses. Besides a hundred chemical compounds that have been isolated, the mai active constituents are flavonoids, alkaloids, phenolic acids, and fatty acids. <i>T. stans</i> exerted		
<i>Keywords:</i> Bignoniaceae Flavonoids Alkaloids Antioxidants Fatty acids	many medicinal benefits, including antidiabetic, anti-inflammatory, anti-cancer, anti- microbial, antioxidant, hepatoprotective, cardioprotective, and nephroprotective properties. However, there is a shortage of in vivo studies, especially adequate dosage and toxicity studies. More studies should be carried out for nutritional data. This review represents a scientific understanding of clinical correlations and applications of phytocompounds from <i>T. stans</i> in protecting and treating many complaints and disorders.		

Implication for health policy/practice/research/medical education:

This work is to present a literature review that categorizes the important phytoconstituents of *Tecoma stans*. Moreover, this review classifies the biological activities of the plant according to the latest published studies. Therefore, it provides beneficial information for future in vivo and clinical studies on *T. stans*.

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Introduction

Tecoma stans (L.) Juss. Ex Kunth (Bignoniaceae) has traditionally been used as a source of medicine contributing to human health and well-being. More than 80% of the world's population uses traditional herbal medicines, especially in the developing world (1). The genetic biodiversity and varieties of traditional medicinal herbs often no longer exist due to growth exploitation, environment-unfriendly harvesting techniques, loss of suitable habitats, and non-monitored dealing with medicinal plants. T. stans is a medicinal herb with many uses for the treatment of diabetes mellitus, gastrointestinal tract complaints, and microbial infections. Moreover, it can be used as a strong diuretic, vermifuge, and tonic. The leaves, bark, and root extracts hold biologically active phytoconstituents; therefore, they are used in traditional folk medicines. The preliminary phytochemical analysis resulted in the presence of many active constituents such

as alkaloids, flavonoids, tannins, quinones, and traces of saponins and amino acids (2).

The generic name of T. stans comes from the word tecomaxochit, which was used by Mexican indigenous peoples to describe plants with tubular flowers, also known as yellow bells, yellow-elder, yellow trumpet bush, trumpet bush, ginger-thomas, esperanza, and tronadora (3). Tecoma is a genus comprising fourteen species of shrubs or small trees in the trumpet vine family, Bignoniaceae. Twelve species are present in the Americas, but the other two species are present in Africa (4). It is distributed throughout South America and India, although its native habitat extends from southern Texas, New Mexico, and Arizona to Bolivia and northern Argentina and from Florida and the Bahamas to Trinidad in the Caribbean. In addition, it is cultivated in tropical and subtropical areas in Africa, Asia, the Pacific Islands, and Australia (3). T. stans survives by fully being exposed

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to sunlight and distributes on roadsides. *T. stans* exists in moderate salt and alkaline soil conditions and propagates by seeds and green cuttings (5). The plant takes about two years to bloom (6).

Morphologically, *T. stans* is about 20-30 ft in height with moderate growth, clustered elongated fruits, yellow flowers, green compound, imparipinnate, lanceolate leaves with a serrate margin of 2 to 5 pairs of leaflets with a larger single terminal leaflet. Leaflets are lanceolate, about 10 cm long, with serrated margins, mid-green above, and softly touched. Flowers are presented in trumpet-shaped clusters at the branches' ends and with 5 rounded lobes, 6 cm long, pale to bright yellow. Fruits are narrow, slightly flattened to pointed capsules, about 20 cm long, with many winged young green seeds turned to pale brown on ripening and maintained in untidy clusters on the tree for months (7).

Some reviews have already been published regarding the constituents and biomedical applications of T. stans. However, some of the published reviews focused on a particular therapeutic use of T. stans, such as its use as an anti-cancer (8) or anti-diabetic (9) remedy, while others covered general information regarding the phytochemistry and the pharmacology of the plant. For example, Anand and Basavaraju (10) categorized the phytoconstituents of T. stans according to the part of the plant from which they were isolated and summarized the findings of the earlier studies about the general medicinal use of the plant. Rahmatullah et al (11) summarized the general uses of T. stans as a member of the family Bignoniaceae. In this review, we categorized the main phytoconstituents isolated from T. stans according to their phytochemical class (such as alkaloids, glycosides, and phenolic compounds). Moreover, we provided a new classification of the most common biological activities of T. stans and summarized most of the previously published studies that were concerned with T. stans' biomedical uses. The information in this review may be a base for upcoming in vivo and clinical research using T. stans.

Taxonomical classification

Domain – Eukaryota

Kingdom - Plantae - plants, Planta, Vegetal, plants

Subkingdom - Viridaeplantae - green plants Infrakingdom - Streptophyta - land plants Phylum - Tracheobionta - Vascular plant Subphylum - Euphyllophytina Super division - Spermatophyta Division - Tracheophyta - vascular plants, tracheophytes Subdivision-Spermatophytina-spermatophytes,seed plants, phanérogames Infradivision - Angiospermae - flowering plants, angiosperms, plantas comflor, angiosperma, plantes à fleurs, angiospermes, plantes à fruits Class- Magnoliopsida Superorder – Asteranae Order - Lamiales Family - Bignoniaceae - bignonias Genus - Tecoma Juss. - trumpetbush Species - Tecoma stans (L.) Juss. ex Kunth - yellow elder,

Vernacular names

Hindi – Piliya/ Pila kaner, English – Yellow bells, Kannada – Koranekelar, Tamil – sonnapatti, Telugu Pachagotla, Bengali – chandaprabha, Marathi – Ghanti ful ².

vellow trumpet flower, trumpet bush, trumpet flower².

Phytochemical constituents

Various compounds that have been recognized in different parts of *T. stans* were classified into alkaloids (Table 1), phenolics (Table 2), flavonoids (Table 3), glycosides (Table 4), carotenoids (Table 5), pentacyclic triterpenoid (Table 6), and miscellaneous compounds (Table 7). All the listed compounds have a significant role in the biomedical impact of the plant.

Biological activities of *Tecoma stans*

There are many studies carried out for recognizing and evaluating the medicinal and biological activities of *T. stans.* The aerial components have been reported to be used in the treatment of gastritis, diarrhea, and stomach problems. Roots were used as a vermifuge, diuretic, tonic, and for the treatment of snake and rat bites, scorpion stings, and syphilis (25). Various phytoconstituents identified and isolated from *T. stans* have exerted many biological properties, including antioxidant, anti-androgenic,



Table 1. Important isolated alkaloids from Tecoma stans

Table 1. Continued

Compound name	Chemical structure	Part of plant/type of extraction
Tecostanine	H N N	Leaves/diethyl ether/NH3 15% (10%) (15)
Tecomanine		Fruits/ethanol (16,17)
7-Hydroxydehydroskytanthine	ОН	Fruits/ethanol (14)
4-Hydroxytecomanine		Fruits/ethanol (14)
Tecostidine	H	Root, leaves, and twigs/methanol or ethanol (18)
Boschniakinic acid		Root, leaves, and twigs/methanol or ethanol (18)
N-Normethyl skytanthine	H	Root, leaves, and twigs/methanol or ethanol (18)
Boschniakine	O H	Root, leaves, and twigs/methanol or ethanol (18) Fruit/aqueous (12)

Table 2. Important isolated phenolics from Tecoma stans

Compound name	Chemical structure	Plant part/type of extract
Ferulic acid	H	Flowers/methanol (13)
	°	Leaves/methanol (16)
	н	
	6 _{~н}	
Chlorogenic acid		Leaves/hydroethanolic (17)
	н н о	Leaves/methanol (16)
	d d d d d d d d d d d d d d d d d d d	
Cinnamic acid	 Н Т	Leaves/hydroethanolic (17)
	°	Leaves/methanol (18)
	нн	
Gallic acid		Leaves/hydroethanolic (17) Leaves/methanol (16)
	н о н	
	o ho	
Caffeic acid	н Н	Leaves/hydroethanolic (17)
	°	Leaves/methanol (16)
	н	
	H	
	 о _{~н}	
Vanillic acid	H	Leaves/hydroethanolic (17)
	°	Leaves/methanol (16)
	о_ _н	
Isoferulic acid	н	Leaves/methanol (17)
(3-Hydroxy-4-methoxy cinnamic acid)		
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2.4.5 Trimothovy cippomic acid	н	Logyos/mothanol (16)
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Table 2. Continued		
Compound name	Chemical structure	Plant part/type of extract
Rosmarinic acid		Leaves/methanol (16)
O-coumaric acid		Leaves/hydroethanolic (17)
Sinapic acids		Leaves/hydroethanolic (18)
Ellagic acid		Leaves/methanol (16)
P-coumaric acid		Leaves/methanol (16)
P-Hydroxybenzoic acid	H H H	Leaves/methanol (16)
Protocatechuic acid		Leaves/methanol (16)

Table 3. Important isolated flavonoids from Tecoma stans

Compound name	Chemical structure	Part of plant
7-Hydroxyflavone	• 	Leaves/methanol (16)
	H.	
Catechin	н _о н 	Leaves/methanol (16)
	H_O	
Naringin	H	Leaves/methanol (16)
	H-O C	
Flavanone	0 	Leaves/hydroethanolic (17)
Luteolin		Leaves/hydroethanolic (17)
	H_	
	U U U U	
	н-б	
Luteolin 7-O-β-D-neohespridoside	он	Fruit/ethanol (12)
	он он он	
Luteolin 7-O-β-D-glucopyranoside	он	Fruit/ethanol (12)
Diosmetin 7-O-β-D-glucopyranoside	H C C	Flower/ethanol (12)
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	U U U U U U U U U U U U U U U U U U U	
Apigenin	н~о о	Leaves/hydroethanolic extract (12)
		-Leaves/methanol (16)
	H	
	H	
	· •	

Table 3. Continued

Compound name	Chemical structure	Part of plant
Chrysoeriol	H O O H	Leaves/hydroethanolic extract (12) -Flowers, methanol (13)
Kaempferol		Leaves/hydroethanolic (17) -Leaves/methanol (16)
Quercetin		Leaves/hydroethanolic (17) -Leaves/methanol (16)
Rutin		Flowers/alcohol (19) -Fruits/ethanol (12) -Leaves/methanol (16)
Quercitrin		Leaves/methanol (16)
Pyrogallol	H ^O H	Leaves /methanol (16)

Table 4. Important isolated glycosides from Tecoma stans



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Table 5. Important isolated carotenoids from Tecoma stans

Compound name	Chemical structure	Plant part
β-carotene	H + H + H + H + H + H + H + H + H + H +	Flowers/ethanol (21)
Zeaxanthin	$\begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & &$	Flowers/aqueous (22)

Table 6. Important pentacyclic triterpenoid compounds from Tecoma stans

Compound name	Chemical structure	Part of plant
Ursolic acid		Leaves/Ethyl acetate (23) Flowers/methanol (13)
3-Oxo-urs-12-en-28-oic acid		Flowers/methanol (13)

Table 7. Important miscellaneous compounds of Tecoma stans

Compound name	Chemical structure	Part of plant	Chemical class
β-Sitosterols		Leaves/ Hydroethanolic (17)	Phyto sterols
Indole	N N N N N N N N N N N N N N N N N N N	Leaves/methanol (24)	Indolic compounds
skatole		Leaves/methanol (24)	Indolic compounds
Anthranilic acid	H N O O	Leaves/methanol (24)	Amino benzoates
Tryptophan		Leaves/methanol (24)	Amino Acids
Tryptamine	H N-H H	Leaves/methanol (24)	Amines

antifungal, lubricant, alpha-reductase inhibitor, immunomodulator, hypocholesterolemic, antihistaminic, antiarthritic, antieczemic, antiacne, protective cancer, and hepatoprotective activities (10). T. stans was also found to exert potential antimicrobial activities against a wide range of microbial strains listed in Table 8. The medicinal activities against bowel diseases such as anti-spasmodic, antidiarrhea, anti-ulcer, and anthelmintic activities are listed in Table 9. Various miscellaneous activities such as inhibiting platelet aggregation, anti-urolithiasis, antiinflammatory, anti-arthritis, antioxidant, antiproliferation, anti-cancer, anti-diabetic, and antidepressant properties are listed in Table 10. Other miscellaneous pharmaceutical applications are listed in Table 11.

Plant safety and toxicity

The safety and toxicity of the plant were studied and concluded that the median acute toxicity (LD50) of *T. stans*

extract was identified to be <5000 mg/kg body weight in mice. Sub-chronic use for 28 days showed significant weight gain, decrease in platelet levels, decrease in white blood cells, and elevation in blood glucose in comparison to the normal. The hydroethanolic extract showed no adverse events on vital organs and was safe in moderate doses (74).

Conclusion

From this review, it can be deduced that *Tecoma stans* (family Bignoniaceae) is rich in various types of active constituents possessing diverse biological properties such as anti-infective, anti-hyperlipidemic, hepatoprotective, cardioprotective, nephroprotective activities. Therefore, it would be important to extensively investigate the phytochemicals and pharmacological activities for future drug discovery and development.

Table 8. Summary of the previous studies about the antimicrobial activities of Tecoma stans

Biological activity	Part used	Type of extract	Name of strain
	Flowers	Ethanol, ethyl acetate, dichloromethane	Escherichia coli, and Enterococcus faecalis (26)
	Branches and leaves	Methanol	Staphylococcus aureus (ATCC# 6538), methicillin-resistant Staphylococcus aureus (MRSA) 10, and MRSA 11 (27)
	Flowers	N-hexane	Streptococcus mutants, harmful bacteria, cause tooth decay, so T. stans could be used in toothpaste and mouthwashes as an anti-oral pathogen (28)
	Leaves, callus	Methanol, chloroform	Staphylococcus aureus, B. subtilis, Escherichia coli, and Pseudomonas aeruginosa (29)
Antibacterial	Flowers	Water	Staphylococcus aureus and Escherichia coli (30)
activities	Leaves	Water, ethanol, and chloroform	Pseudomonas aeruginosa (31)
	Flowers	Ethanolic	B. subtilis, E. coli, S. aureus, P. mirabilis, and K. pneumoniae (32)
	Plant materials	Chloroform, butanol, and ethyl acetate	E. coli, P. aeruginosa, and S. aureus (33)
	Flowers	Methanolic	E. coli, Entero bacterium, and bacillus creases. Gram-negative strains were more susceptible to raw materials than gram-positive strains (34)
	Plant parts	Ethanol, methanol, and water extracts	E. coli, Klebsiella pneumonia, Clavibacter michiganensis sub sp. Michiganensis, Xanthomonas axonopodis pv. malvacearum, Pseudomonas fluorescens, Pseudomonas aeruginosa, and Staphylococcus aureus (35)
	Flowers	Ethanolic	Penicillium spp (32)
	Leaves	Water, ethanol, chloroform	Aspergillus niger (31)
Antifungal	Plant materials	N-hexane, ethyl acetate	Fusarium solani and Aspergillus niger (33)
activities	Plant parts	Ethanol, methanol, and water extracts	All species of Aspergillus and Alternaria (35)
	Leaves	Petroleum ether, chloroform, ethanol	Candida albicans, Cryptococcus neoformans, and Microsporum gypseum (38)
	Root	Dichloromethane and methanol	Saccharomyces and Candida Albicans (37)
Antiviral Activity	Trunks, leaves	Ethanol	Zika virus (38)

Table 9. Summary of the previous studies of bowel biological activities of Tecoma stans

Biological activity	Part used	Extraction	Summary of the studies
Antispasmodic activity	Leaves	Hydroalcoholic	The antispasmodic effect of the extract may be due to the involvement of calcium channels, without interfering with opioid receptors, β-adrenoceptors, and potassium channels (39).
Antidiarrheal	Flowers	Ethanol	Antidiarrheal-dependent action was significantly identified. There was a potential decrease in fecal fluctuations frequency (40).
Antiulcer	Leaves	Methanol	Flavonoid contents were responsible for the significant effective protection of gastric mucosa against aspirin-induced ulcers at all doses (41)
Activity against inflammatory bowel disease	Flower	Ethanol	The extract exerted anti-inflammatory and antioxidant properties that significantly ameliorate colitis (42).
Anthelmintic activity	Leaves	Methanol	T. stans exerted molluscicidal activities on B. Alexandrina snails, thus it could be an effective, cheap, and environmentally safe agent to control the spread of schistosomiasis (43).
	Flower, root	Ethanol	T. stans was proven to treat helminthiasis. The flower and roots of T. stans decoction are used for intestinal worms. Leaves also had anthelmintic activity (44).

Table 10. Summary of the previous studies of miscellaneous activities of Tecoma stans

Biological activity	Part used	Type of extract	Summary
Inhibition of induced- platelets aggregation	Leaves	Aqueous	Extracts of <i>T. stans</i> exerted a protective function against the proliferation of rabbit artery skin. <i>T. stans</i> significantly inhibited capillary permeability in rabbits in a concentration-dependent manner (45).
Nephroprotective activities	Flowers	Ethyl acetate	The extract exerted protective activity in albino rats against gentamicin-induced nephrotoxicity (46).
	Leaves	Aqueous	The extract exerted strong anti-aggregate activities with thrombin-induced aggregated human platelets (47).
Cardioprotective activity	Leaves	Ethyl acetate, methylene chloride (fractions)	The ethyl acetate fraction showed a maximum reduction of triglyceride, very low-density lipoprotein, and atherogenic index. The methylene chloride fraction riches, in alkaloids, reduced the low-density lipoprotein levels, while the flavonoid fraction (rutin rich) raised the high-density lipoprotein levels (48).
	Flowers	Ethanol	Polyphenolic constituents such as (flavonoids and tannins) showed effective antioxidant properties. Moreover, the extract stopped the deviation of reduced glutathione, superoxide dismutase, and catalase levels in a dose-dependent manner (49).
	Flower	Hydro ethanol	The extract inhibited the elevation in serum cholesterol and triglyceride levels (50).
Hepatoprotective activity	Leaves	Hydroethanolic solution	The extract exerted hepatoprotective activities against liver damage induced by carbon tetrachloride and acetaminophen caused in rats (51).
Antidepressant activities	Flowers	Methanolic and aqueous	These extracts contained Flavonoids that exert antidepressant activity (52).
Wound healing activity	Flower	Ethanol	Ointment of the extract was identified to heal all wound models with significant improvement of all parameters (53).
	Bark	Petroleum ether, chloroform, and methanol	The methanolic extract resulted in wound healing improvement due to the presence of phytocomponents likes, phenols, flavonoids phytosterols, triterpene, glycosides, saponins, and tannins. The antimicrobial effects of <i>T. stans</i> play a role in wound healing (54).
	Bark, leaves, flowers	Ethanol	The leaf and flower worked better than the bark. The extract of the leaves and flowers of <i>T. stan</i> was found to be 100% more effective against inflammation than the common drug ibuprofen (55).
Anti-inflammatory activity	Flowers	Methanol	The extract showed marked anti-nociceptive activity. This may be due to the presence of flavonoids which may play a role in the inhibition of prostaglandin synthesis (56).
	Leaves	Alcohol and aqueous	Both extracts exhibited dose-dependent activity. Alcohol extract showed the highest reduction of inflammation after 24 hrs., because of the presence of higher phenolic and flavonoid content (57).
Anti-arthritic activity	Stem	Methanol	The extract potentially reduced protein denaturation and decreased the inflammation volume. The anti-arthritic activity may be due to the flavonoids, alkaloids, steroids, and glycosides in the extract (58).
	Leaves	Alcohol, water, Petroleum ether, chloroform, methanol.	Alcohol, Water successive methanol extracts exhibited significant Antiarthritic activity (59).

Table 10. Continued

Biological activity	Part used	Type of extract	Summary
Antioxidant activities	Leaves, branches	Methanol	The crude extract exhibited significant activity of 80% maximum scavenging potential (27).
	Leaves, flowers	Methanol	The higher concentrations showed higher antioxidant activities than the standard ascorbic acid (60).
	Root, bark	Ethanol	T. stans was most efficient in a concentration-dependent manner in comparison to different plants by using a 2,2-diphenyl-1-picryl-hydrazyl-hydrate (DPPH) radical, superoxide anion radical, and nitric oxide radical scavenging capabilities for antioxidants (61).
	Leaves, branches	Ethyl acetate, chloroform, methanol	T. stans leaves and branches ethyl acetate fraction exerted the best total antioxidant activity followed by chloroform fraction of leaves extracts, methanol extract from leaves, and then methanol extract of branches (62).
	Plant Parts	Methanol, ethanol, water Plant parts	All solvent fractions showed strong radical scavenging activity from a ferric-reducing antioxidant power assay and DPPH (35).
Anti-proliferative, cytotoxic, and anticancer activities	Branches, leaves	Methanol	Anticancer dose-dependent activity against the rhabdomyosarcoma cell line was exhibited by the methanolic extract (27).
	Leaves, flowers	Methanol	Used as a remedy for lung cancer and can be an effective ingredient in cancer drug recipes (60).
	Leaves	Ethanolic	The extract exhibited anticancer activity against breast cancer cell line (MCF-7) with different concentrations (63).
	Leaves	Water	Cytotoxic effects of T. stans were concentration, and time-dependent against a (HepG2) cell line in the presence and absence of fetal bovine serum (64, 65).
Antidiabetic activity	Stem	Ethanol	The extract could prevent subsequent diabetic complications as diabetic nephropathy and retinopathy (12).
	Flowers	Ethanol	T. stans showed significant anti-apoptotic potential. Significantly reduced myocyte loss augmented the cardiac activity in rats with diabetes (66).
	Flowers	Alcohol and water	The extracts reduced blood glucose (67).
	Leaves	Water and alcohol	Both T. stans (L.) Kunth cv. Nalgonda 1 and T. stans (L.) Kunth cv. Warangal 1 extracts could lower blood glucose levels after a reduction in the breakdown of carbohydrates to light sugars (68).

Table 11. Summary of the previous studies of pharmaceutical activities of Tecoma stans

Pharmaceutical activities	Part used	Type of extract	Summary
As a natural dye	Flowers	Ethanol, and ethanol, a water mixture	Flowers of <i>T. stans</i> could be a potential natural source of yellow colorant (69). Moreover, they were used as natural dye sensitizers in dye solar cells (70).
As reducing/capping agent	Leaves	Water	The extract was used as a reducing agent in the synthesis of the reduced graphene oxide which was used for the removal of Ni from water (71).
		Water	The extract was used as a reducing agent and capping agent in the synthesis and stabilization of silver nanoparticles (72).
As an insecticide agent	Leaves	Ethanol	Leaf extract showed larvicidal activity against the mosquito species Culex pipiens L. (Diptera: Culicidae) (73).

Author's contributions

NEW and MF conceptualized the study. AK and DAF prepared the manuscript. All authors reviewed, confirmed, and approved the final version of the manuscript.

Conflict of interests

No potential conflict of interest was reported by the authors.

Ethical considerations

Ethical issues (including plagiarism, data fabrication, double publication, etc.) have been completely observed by the authors.

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References

- World Health Organization (WHO). WHO Guidelines on Good Agricultural and Collection Practices (GACP) for Medicinal Plants. Geneva: WHO, 2003. p. 72.
- 2. Singh A, Nagori BP, Mathur K. *Tecoma stans*: an important medicinal plant. Int J Pharm Erud. 2013;3(2):13-21.
- Anand M, Basavaraju R. GC analysis of different parts of *Tecoma stans* (L.) Juss. ex Kunth for fathy acid composition. Curr Bot. 2021;12:16-21. doi: 10.25081/cb.2021.v12.6605.
- Jedidi B, Mokbli S, Sbihi HM, Nehdi IA, Romdhani-Younes M, Al-Resayes SI. Effect of extraction solvents on fatty acid composition and physicochemical properties of *Tecoma stans* seed oils. J King Saud Univ Sci. 2020;32(4):2468-73. doi: 10.1016/j.jksus.2020.03.044.
- Orwa C, Mutua A, Kindt R, Jamnadass R, Anthony S. Agroforestry Database: A Tree Reference and Selection Guide Version 4.0. Kenya: World Agroforestry Centre; 2009. p. 1-5.
- White RC. Elsevier's Dictionary of Plant Names of North America, Including Mexico. 1st ed. Amsterdam: Elsevier; 2003. p. 193.
- Abisha Vince Jeo VS, Justin Raj S. Therapeutic properties and applications of *Tecoma stans* Linn. Int J Pharm Sci Rev Res. 2020;63(1):111-5.
- Jaikumar B, Jasmine R. A review on a few medicinal plants possessing anticancer activity against human breast cancer. Int J PharmTech Res. 2016;9(3):333-65.
- Gupta A, Behl T. Proposed mechanism of *Tecoma* stans in diabetes-associated complications. Nat Prod J. 2021;11(2):127-39. doi: 10.2174/221031551066619122411 4311.
- 10. Anand M, Basavaraju R. A review on phytochemistry and pharmacological uses of *Tecoma stans* (L.) Juss. ex Kunth. J Ethnopharmacol. 2021;265:113270. doi: 10.1016/j. jep.2020.113270.
- Rahmatullah M, Samarrai W, Jahan R, Rahman S, Sharmin N, Miajee EU, et al. An ethnomedicinal, pharmacological and phytochemical review of some Bignoniaceae family plants and a description of Bignoniaceae plants in folk medicinal uses in Bangladesh. Adv Nat Appl Sci. 2010;4(3):236-53.
- 12. Marzouk M, Gamal-Eldeen A, Mohamed M, El-Sayed

M. Anti-proliferative and antioxidant constituents from *Tecoma stans*. Z Naturforsch C J Biosci. 2006;61(11-12):783-91.

- Ha KN, Nguyen TVA, Mai DT, Tran NMA, Nguyen NH, Vo GV, et al. Alpha-glucosidase inhibitors from *Nervilia* concolor, Tecoma stans, and Bouea macrophylla. Saudi J Biol Sci. 2022;29(2):1029-42. doi: 10.1016/j.sjbs.2021.09.070.
- Lins AP, Felicio JDA. Monoterpene alkaloids from *Tecoma stans*. Phytochemistry. 1993;34(3):876-8. doi: 10.1016/0031-9422(93)85381-Z.
- Costantino L, Lins AP, Barlocco D, Celotti F, el-Abady SA, Brunetti T, et al. Characterization and pharmacological actions of tecostanine, an alkaloid of *Tecoma stans*. Pharmazie. 2003;58(2):140-2. doi: 10.1002/chin.200324180.
- Taher MA, Dawood DH, Sanad MI, Hassan RA. Searching for anti-hyperglycemic phytomolecules of *Tecoma stans*. Eur J Chem. 2016;7(4):397-404. doi: 10.5155/ eurjchem.7.4.397-404.1478.
- Ramirez G, Zamilpa A, Zavala M, Perez J, Morales D, Tortoriello J. Chrysoeriol and other polyphenols from *Tecoma stans* with lipase inhibitory activity. J Ethnopharmacol. 2016;185:1-8. doi: 10.1016/j. jep.2016.03.014.
- Dickinson EM, Jones G. Pyrindane alkaloids from *Tecoma* stans. Tetrahedron. 1969;25(7):1523-9. doi: 10.1016/s0040-4020(01)82725-3.
- Rajendran A. Isolation, characterization, pharmacological and corrosion inhibition studies of flavonoids obtained from *Nerium oleander* and *Tecoma stans*. Int J PharmTech Res. 2011;3(2):1005-13.
- Bianco A, Massa M, Oguakwa JU, Passacantilli P. 5-deoxystansioside, an iridoid glucoside from *Tecoma stans*. Phytochemistry. 1981;20(8):1871-2. doi: 10.1016/0031-9422(81)84024-1.
- Rajamurugan R, Thirunavukkarasu C, Sakthivel V, Sivashanmugam M, Raghavan CM. Phytochemical screening, antioxidant and antimicrobial activities of ethanolic extract of *Tecoma stans* flowers. Int J Pharma Bio Sci. 2013;4(2):124-30.
- Singh S, Maurya IC, Sharma S, Srivastava P, Bahadur L. Effect of dye extraction solvent on the photovoltaic performance of *Tecoma stans*-sensitized solar cells. J Electron Mater. 2020 ;49(7):4355-4363.
- 23. Kedar KA, Chaudhari SR, Rao AS. Development of a densitometric high-performance thin-layer chromatographic method for the quantitative analysis of ursolic acid in the leaves of species of genus *Tecoma* and *Tabebuia* of Bignoniaceae family. Int J Pharm Pharm Sci. 2017;9(2):109-13. doi: 10.22159/ijpps.2017v9i2.15802.
- Kunapuli SP, Vaidyanathan CS. Indolic compounds in the leaves of *Tecoma stans*. Phytochemistry. 1984;23(8):1826-7. doi: 10.1016/s0031-9422(00)83513-x.
- Sharma PC, Yelne M, Dennis TJ. Database on medicinal plants used in ayurveda. New Delhi: Central Council for Research in Ayurveda and Siddha (India). 2000. p. 378-83.
- Gonçalves TPR, Parreira AG, dos Santos Zanuncio VS, de Souza Farias K, da Silva DB, dos Santos Lima LAR. Antibacterial and antioxidant properties of flowers from *Tecoma stans* (L.) Juss. ex Kunth (Bignoniaceae). S Afr J Bot. 2022;144:156-65. doi: 10.1016/j.sajb.2021.08.028.

- Tariq H, Rafi M, Amirzada MI, Muhammad SA, Yameen MA, Mannan A, et al. Photodynamic cytotoxic and antibacterial evaluation of *Tecoma stans* and *Narcissus tazetta* mediated silver nanoparticles. Arab J Chem. 2022;15(3):103652. doi: 10.1016/j.arabjc.2021.103652.
- Mohammed HA, Abdel-Aziz MM, Hegazy MM. Antioral pathogens of *Tecoma stans* (L.) and *Cassia javanica* (L.) flower volatile oils in comparison with chlorhexidine in accordance with their folk medicinal uses. Medicina (Kaunas). 2019;55(6):301. doi: 10.3390/medicina55060301.
- 29. Khatak S, Malik DK, Dahiya R. *Tecoma stans*: a noxious weed put to beneficial use. Int J Chem Stud. 2019;7(3):296-9.
- Hariram M, Vivekanandhan S, Ganesan V, Muthuramkumar S, Rodriguez-uribe A, Mohanty AK, et al. *Tecoma stans* flower extract assisted biogenic synthesis of functional Ag-Talc nanostructures for antimicrobial applications. Bioresour Technol Rep. 2019;7:100298. doi: 10.1016/j. biteb.2019.100298.
- 31. Dewangan N, Satpathy S, Shrivastava AK, Shrivastava RA. In vitro evaluation of antimicrobial activity of *Tecoma stans* and *Vitex negundo*. Indian J Sci Res. 2017;13(2):248-53.
- Khan AS. Trees with antimicrobial activities. In: Khan AS, ed. Medicinally Important Trees. Cham: Springer; 2017. p. 85-108. doi: 10.1007/978-3-319-56777-8_4.
- 33. Javid T, Adnan M, Tariq A, Akhtar B, Ullah R, Abd El Salam NM. Antimicrobial activity of three medicinal plants (*Artemisia indica, Medicago falcate* and *Tecoma stans*). Afr J Tradit Complement Altern Med. 2015;12(3):91-6. doi: 10.4314/ajtcam.v12i3.11.
- Mamone L, Di Venosa G, Gándara L, Sáenz D, Vallecorsa P, Schickinger S, et al. Photodynamic inactivation of Gram-positive bacteria employing natural resources. J Photochem Photobiol B. 2014;133:80-9. doi: 10.1016/j. jphotobiol.2014.03.003.
- Govindappa M, Sadananda TS, Channabasava R, Jeevitha MK, Pooja KS, Raghavendra VB. Antimicrobial, antioxidant activity and phytochemical screening of *Tecoma stans* (L.) Juss. ex Kunth. J Phytol. 2011;3(3):68-76.
- Indra Gandhi M, Ramesh S. Antifungal and haemolytic activities of organic extracts of *Tecoma stans* (Bignoniaceae). J Ecobiotechnol. 2010;2(2):26-32.
- Kumar VP, Chauhan NS, Padh H, Rajani M. Search for antibacterial and antifungal agents from selected Indian medicinal plants. J Ethnopharmacol. 2006;107(2):182-8. doi: 10.1016/j.jep.2006.03.013.
- Reis ACC, Silva BM, de Moura HMM, Pereira GR, Brandão GC. Anti-Zika virus activity and chemical characterization by ultra-high performance liquid chromatography (UPLC-DAD-UV-MS) of ethanol extracts in *Tecoma* species. BMC Complement Med Ther. 2020;20(1):246. doi: 10.1186/ s12906-020-03040-0.
- Gharib Naseri MK, Asadi Moghaddam M, Bahadoram S. Antispasmodic effect of *Tecoma stans* (L.) Juss leaf extract on rat ileum. Daru. 2007;15(3):123-8.
- 40. Kameshwaran S, Jothimanivannan C, Senthilkumar R, Thenmozhi S, Sundaraganapathy R, Dhanalakshmi M. Acute toxicity study and fecal dropping capability of ethanolic extract of *Tecoma stans* in albino rats. Pharmacologia. 2013;4(7):464-8. doi: 10.17311/ pharmacologia.2013.464.468.

- Shanmukha I, Vijay kumar M, Ramachandra Setty S. Protective effect of *Tecoma stans* leaf extract on experimentally induced gastric ulcers in rats. Int J Drug Dev Res. 2013;5(3):231-6.
- 42. Kameshwaran S, Senthil Kumar R, Thenmozhi S, Dhanalakshmi M, Vasuki K, Manjuladevi K. Effect of ethanolic extract of flowers of *Tecoma stans* on acetic acid induced colitis in albino rats. Asian J Pharm Sci Technol. 2014;4(2):77-82.
- 43. Ibrahim AM, Ahmed AK, Hammam OA, Abdel-Ghaffar F. Immunotoxical, neurotoxical, histopathological and immunohistopathological alterations of *Nerium oleander* and *Tecoma stans* methanolic extract on *Biomphalaria alexandrina* snails. Acta Trop. 2022;230:106405. doi: 10.1016/j.actatropica.2022.106405.
- Kumanan R, Sridhar C, Jayaveera KN, Sudha S, Rubesh Kumar S, Duganath N. Comparative study of anthelmintic activity of different leaf extracts of *Tecoma stans* (L.) on adult Indian earthworms. Int J Pharm Clin Res. 2010;2(2):63-5.
- 45. Gutiérrez RM, Solís RV. Effect on capillary permeability in rabbits of *Acalypha langinia*, *Buddleia scordioides*, *Hylocereus undatus*, *Tecoma stans* and *Astianthus viminalis*. Pharmacologyonline. 2006;1:113-9.
- 46. Raju S, Kavimani S, Uma Maheshwara rao V, Sreeramulu Reddy K, Vasanth Kumar G. Floral extract of *Tecoma stans*: a potent inhibitor of gentamicin-induced nephrotoxicity in vivo. Asian Pac J Trop Med. 2011;4(9):680-5. doi: 10.1016/ S1995-7645(11)60173-9.
- Villar R, Calleja JM, Morales C, Cáceres A. Screening of 17 Guatemalan medicinal plants for platelet antiaggregant activity. Phytother Res. 1997;11(6):441-5. doi: 10.1002/ (sici)1099-1573(199709)11:6<441::aid-ptr126>3.0.co;2-t.
- Lins AP, Felicio JDA. Monoterpene alkaloids from *Tecoma stans*. Phytochemistry. 1993;34(3):876-8. doi: 10.1016/0031-9422(93)85381-z.
- 49. Ittagi S, Merugumolu VK, Siddamsetty RS. Cardioprotective effect of hydroalcoholic extract of *Tecoma stans* flowers against isoproterenol induced myocardial infarction in rats. Asian Pac J Trop Dis. 2014;4(Suppl 1):S378-S84. doi: 10.1016/s2222-1808(14)60474-6.
- Giri RK, Kanungoa SK, Tripathib NK. Lipid lowering activity of the hydro-alcoholic extract of *Tecoma stans* L. flowers in hyperlipidemic models of Wistar albino rats. Der Pharm Lett. 2012;4(5):1386-9.
- Larbie C, Emikpe BO, Akpor SA, Adams E, Adjei CO, Oyagbemi AA, et al. Ameliorative effect of extract of *Tecoma stans* (L.) Juss. ex Kunth leaves against CCl4-and acetaminophen—induced liver damage in rats. Adv Tradit Med. 2020;20(4):555-62. doi: 10.1007/s13596-020-00465-3.
- 52. Khare CP. Indian Medicinal Plants: An Illustrated Dictionary. Springer; 2007. p. 649-50.
- 53. Kameshwaran S, Senthilkumar R, Thenmozhi S, Dhanalakshmi M. Wound healing potential of ethanolic extract of *Tecoma stans* flowers in rats. Pharmacologia. 2014;5(6):215-21.
- 54. Das C, Dash S, Sahoo DC, Mohanty A. Evaluation of methanolic bark extract of *Tecoma stans* Linn, for wound healing in albino rats. Int J Pharm Technol. 2010;2(3):735-42.
- 55. Swarna SK, Nivedhitha MS, Vishnu PriyaV, Gayathri R, Selvaraj J, Madhan K, et al. Comparative evaluation of

anti-inflammatory potential of ethanolic extract of leaf, bark and flower of *Tecoma stans* with ibuprofen-an in vitro analysis. Pharmacogn J. 2019;11(5):1088-92. doi: 10.5530/ pj.2019.11.170.

- Kameshwaran S, Suresh V, Arunachalam G, Frank PR, Manikandan V. Evaluation of antinociceptive and antiinflammatory potential of flower extract *Tecoma stans*. Indian J Pharmacol. 2012;44(4):543-4. doi: 10.4103/0253-7613.99352.
- Prasanna VL, Lakshman K, Hegde MM, Bhat V. Antinociceptive and anti-inflammatory activity of *Tecoma stans* leaf extracts. Indian J Res Pharm Biotechnol. 2013;1(2):156-60.
- Santhosh E, Gangaraju M, Vivek KT. Evaluation of antiarthritic potentials of methanolic stem extract of *Tecoma stans* Linn in Wistar albino rat. J Rheumatol Arthritic Dis. 2018;3(1):1-6.
- 59. Prajapati D, Patel N. In vitro anti-arthritic activity of *Tecoma stans* (Linn.) leaves. Alger J Nat Prod. 2015;3(2):153-8.
- Robinson JP, Suriya K, Subbaiya R, Ponmurugan P. Antioxidant and cytotoxic activity of *Tecoma stans* against lung cancer cell line (A549). Braz J Pharm Sci. 2017;53(3):e00204. doi: 10.1590/s2175-97902017000300204.
- Chinaka NC, Monago-Ighorodje CC, Chuku LC, Nzom UG, Ohagwu US, Okparoka CC. Antioxidant and free radical scavenging activity of ethanolic extracts of *Tecoma stans*, *Glyphaea brevis*, *Garcinia kola*, *Zanthoxylum macrophylla* and *Gongronema latifolium* root bark. World J Pharm Res. 2017;6(4):248-57. doi: 10.20959/wjpr20174-7956.
- 62. Salem MZ, Gohar YM, Camacho L, El-Shanhorey NA, Salem AZM. Antioxidant and antibacterial activities of leaves and branches extracts of *Tecoma stans* (L.) Juss. ex Kunth against nine species of pathogenic bacteria. Afr J Microbiol Res. 2013;7(5):418-26. doi: 10.5897/ajmr12.2274.
- 63. Thirumal M, Kishore G, Prithika R, Das S, Nithya G. In vitro anticancer activity of *Tecoma stans* (L.) ethanolic leaf extract on human breast cancer cell line (MCF-7). Int J Pharm Chem Biol Sci. 2012;2(4):488-93.
- Zhu J, Viñas R, Smith EE. In vitro evaluation of human liver cancer cells and the potential cytotoxicity of *Tecoma* stans (Bignoniaceae) and *Brickellia cavanillesi* (Asteraceae) both single and in combination. Toxicol Environ Chem. 2008;90(4):801-8. doi: 10.1080/02772240701740387.
- 65. Gaitán I, Paz AM, Zacchino SA, Tamayo G, Giménez A, Pinzón R, et al. Subcutaneous antifungal screening of Latin

American plant extracts against *Sporothrix schenckii* and *Fonsecaea pedrosoi*. Pharm Biol. 2011;49(9):907-19. doi: 10.3109/13880209.2011.555916.

- 66. Sugavanam K, Senthilkumar R, Shanmugam T. Ameliorative effect of *Tecoma stans* extract on diabetic cardiomyopathy against streptozotocin-induced diabetes in Wistar rats. J Pharm Pharmacol. 2013;1:55-62.
- 67. Dhaked U, Gupta V, Singh DP, Nama G. Antidiabetic activity of *Tecoma stans* flower. Pharmacologyonline. 2011;1:553-8.
- Rao KN, Swarna K, Banji D, Sandhya S. Establishment of two varieties in *Tecoma stans* of Indian origin pharmacognostically and pharmacologically. J Phytol. 2010;2(8):92-102.
- 69. Arunkumar P, and Yogamoorthi A. Isolation, application and biochemical characterization of colour component from *Tecoma stans*: a new cost effective and eco-friendly source of natural dye. Int J Nat Prod Res. 2014;4(1):9-11.
- Singh S, Maurya IC, Sharma S, Srivastava P, Bahadur L. Effect of dye extraction solvent on the photovoltaic performance of *Tecoma stans*-sensitized solar cells. J Electron Mater. 2020;49(7):4355-63. doi: 10.1007/s11664-020-08154-2.
- Mahmoud AED, Hosny M, El-Maghrabi N, Fawzy M. Facile synthesis of reduced graphene oxide by *Tecoma stans* extracts for efficient removal of Ni(II) from water: batch experiments and response surface methodology. Sustain Environ Res. 2022;32(1):22. doi: 10.1186/s42834-022-00131-0.
- Vivekanandhan S, Venkateswarlu M, Carnahan D, Misra M, Mohanty AK, Satyanarayana N. Functionalization of single-walled carbon nanotubes with silver nanoparticles using *Tecoma stans* leaf extract. Physica E Low Dimens Syst Nanostruct. 2012;44(7):1725-9. doi: 10.1016/j. physe.2011.10.013.
- 73. Hafsi N, Hamaidia K, Soltani N. Chemical screening, insecticidal and reprotoxic activities of *Tecoma stans* ethanolic leaf extract against the vector mosquito *Culex pipiens*. Physiol Entomol. 2022;47(3):176-87. doi: https:// doi.org/10.1111/phen.12386.
- Larbie C, Owusu Nyarkoh C, Owusu Adjei C. Phytochemical and safety evaluation of hydroethanolic leaf extract of *Tecoma stans* (L.) Juss. ex Kunth. Evid Based Complement Alternat Med. 2019;2019:7417624. doi: 10.1155/2019/7417624.