

http://www.herbmedpharmacol.com



# Journal of Herbmed Pharmacology

# **Rethinking the basic action modes of herbal medicine and pondering classical standardization**



Azis Saifudin<sup>1\*®</sup>, Muh. Akbar Bahar<sup>2®</sup>, Muh. Haqqi Hidayatullah<sup>1®</sup>, Hisayoshi Norimoto<sup>3®</sup>, Yasuhiro Tezuka<sup>4®</sup>, Ken Tanaka<sup>5®</sup>

<sup>1</sup>Faculty of Pharmacy, Universitas Muhammadiyah Surakarta, Pabelan, KTS Solo, Jawa Tengah 57102, Indonesia
 <sup>2</sup>Department of Pharmacy, Faculty of Pharmacy, Universitas Hasanuddin, Makassar, South Sulawesi, 90245, Indonesia
 <sup>3</sup>Pura Pharm Japan Corporation, Room 406, 4th FL., JOHO BLDG., 527 Takata, Toyama, 930-0866, Japan
 <sup>4</sup>Department of Pharmacognosy, College of Pharmaceutical Science, Ritsumeikan University, 1-1-1 Nojihigashi, Kusatsu, Shiga 525-8577, Japan

<sup>5</sup>Faculty of Pharmaceutical Sciences, Hokuriku University, Ho-3, Kanagawa machi, Kanazawa 920-1181, Japan

| ARTICLEINFO  | A B S T R A C T   |  |  |  |
|--|---|--|--|--|
| Article Type:<br>Review  | Over the past two decades, the secondary metabolite platform has determined the scientific direction of herbal medicines, while plant sources have been assumed to be the object of   |  |  |  |
| <i>Article History:</i><br>Received: 22 September 2023<br>Accepted: 16 January 2024  | lead discoveries through bioassay-guided fractionation efforts. Nonetheless, the majority of purification programs have resulted in fractions and pure compounds with much lower efficacy than their parent extracts. It is then assumed that co-working action modes among chemical constituents occur in the herbal preparations. Primary metabolites (polysaccharides  |  |  |  |
| <i>Keywords:</i><br>Synergistic action<br>Additive action<br>Complementary medicine<br>Herbal preparations<br>Bioassay guided-fractionation<br>Secondary metabolites | peptides, and fatty acids) and mineral groups, on the other hand, have been neglected in<br>the herbal effect contributions. This review aims to understand the interplay of secondary<br>metabolites in herbal preparations, particularly how they interact with primary metabolites<br>and mineral groups. Thus, by adhering to classical methods, it is possible to address certain<br>aspects that modern standardization lacks, thereby facilitating a more comprehensive<br>approach to these issues. |  |  |  |

#### *Implication for health policy/practice/research/medical education:*

The study demonstrates that multi-component herbal preparation paradigms should be revisited to prepare health and medical students for future multidisciplinary teamwork. In the industrial sector, integrating coworking action in herbal formulations is essential to establish a solid foundation for advancing herbal product development, particularly for comprehensive herbal component standardization.

*Please cite this paper as:* Saifudin A, Bahar MA, Hidayatullah MH, Norimoto H, Tezuka Y, Tanaka K. Rethinking the basic action modes of herbal medicine and pondering classical standardization. J Herbmed Pharmacol. 2024;13(2):163-175. doi: 10.34172/jhp.2024.48269.

#### Introduction

Throughout the course of history, herbal medicines have garnered recognition for their esteemed status as substances that promote and maintain human health throughout many societies. Indeed, the initiation of the drug discovery process has been the isolation of natural chemicals obtained from plants. Notable examples include the extraction of morphine for its pain-relieving properties. Subsequently, this approach has led to the discovery of various drugs with anticancer properties such as taxol, camptothecin, vincristine, and vinblastine. Additionally, cardiotonic digoxin, anti-malarial agents like artemisinin and quinines, and anti-Alzheimer's medications such as galanthamine have also been derived from natural compounds (1,2). However, there has been a noticeable cessation in the development of novel pharmaceuticals derived from plants during the last twenty years. Subsequently, a significant proportion of natural drug discovery initiatives have transitioned towards using microbial or marine screening sources (3-5). Conversely, in the contemporary era of evidencebased medicine, herbal extract preparations have been categorized as complementary alternative medicines, specifically designed as preventative and promotional

#### Saifudin et al

agents, owing to their uncertain outcomes. Some official authorities have provided herbal pharmacopoeias with comprehensive monograph outlines that include information on botanicals, secondary metabolite profiling, and contaminant threshold requirements, which align with the modern chemical-pharmacological paradigm. Since the uncertainty of outcomes has been mitigated through qualitative and quantitative analysis of chemical constituents, the main mission of these official documents is to focus on the safety of herbal preparations. The idea that certain isolated compounds from plants exhibit biological effects and have already been used in clinical therapy has led to the modernization of herbal medicine, especially in the use of the term specific standardization (6,7).

A number of major compounds in plant extracts could be the active principal constituents, such as hydroxycinnamic acid derivatives in Sarcocaulon marlothii stem (8), andrographolide in Andrographis paniculata leaf (9), 1'-acetoxychavicol acetate in Alpinia galanga rhizome (10), linalool in Coriandrum sativum fruit (11), xylaranic acid in Xylaria primorskensis (12), and hyperforin in Hypericum perforatum herb (13). The vast majority of perpetual bioassay-guided fractionation work has been aimed at discovering active compounds from crude materials. Technically, bioassay-guided fractionation is used to link drug discovery screening with biological responses mediated by an instrument analysis. It is done step by step from crude extract to purified materials (14). Unfortunately, numerous studies employing bioassayguided fractionation have yielded bioactivity effects that are statistically insignificant for the isolated compounds when compared to their original extracts (15-17). In certain cases, these isolated compounds have even demonstrated lower effects (18-20), or no effects at all (21,22). Moreover, with regards to effectiveness, the primary active chemicals are frequently discovered in limited concentrations. There is not always a clear link between the amount of these compounds in the active parent extracts and their biological effects (23). The aforementioned concerns have been brought to attention through many findings on bioassay-guided fractionation, indicating the potential limitations of depending exclusively on certain chemicals as the main active metabolite (24-27). This review aimed to elucidate the current mainstream standardization of herbal medicinal preparations that may have deviated from the basic pharmacological theory. Further, it provides an evaluation of certain classical standardization methods as an alternative to address the limitations of some claimed comprehensive standardization methods.

# Coworking mode of actions

#### Synergistic interaction

The first form of coworking mode of action is synergistic interaction, wherein the combination of two natural

compounds yields effects greater than the cumulative responses elicited by each substance when administered individually (the general formula of 1+1 > 2). The concept of pure synergism pertains to the phenomenon in which the presence of a non-active metabolite contributes to the augmentation or modulation of the effects of an active molecule. As recently revealed by Vidar et al (28), berberine antibacterial effect was enhanced by the presence of piperine. On the contrary, piperine itself did not have any antibacterial activity. Previously, Bilia et al (29) revealed that the antiplasmodial effect of artemisinin was enhanced by the presence of quercetin in certain ratios. Nevertheless, in vivo or clinical synergistic data should also be considered since Elfawal et al (30) and Cai et al (31) reported that some flavonoids inhibited P450 cytochromic metabolism of artemisinin and increased blood artemisinin concentration. Thus, in the context of non-pure synergy, the presence of a less active molecule enhances the effects of an active metabolite. The aforementioned phenomena have been elucidated by the utilization of in vitro antioxidant assays conducted on some polyphenols (32-34).

#### Additive interaction

The second proposed coworking modes is characterized by additive interaction, in which some compounds within mixtures have a shared action target. The observed quantitative effects arise from the aggregation of the individual effects of the compounds when they are given independently (follows the basic formula of 1 + 1 with approximate result of 2).

However, the additive mode is typically avoided in the clinical settings because of the possibility of unfavorable adverse outcomes (35,36). The published data on the interaction among natural compounds through pure additive mechanism, resulting from the combination among partial agonists, is quite scarce. Therefore, the explanation for additive mechanism could be approached by referring to the in vitro assay models such as antioxidant assays. As described by Heo et al in their endeavor to unveil the synergistic effects among phenolic catechin, chlorogenic acid, cyanidin, cyanidin 3-glucoside, cyanidin 3-rutinoside, epicatechin, peonidin, peonidin 3-glucoside, quercetin, quercetin 3-glucoside, quercetin 3-galactoside, and quercetin 3-rutinoside through ABTS (2,20-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) assay method, they reported that the individual capacity of these phenolic compounds was similar to the summation of their combination effects (37). Hence, there was additive instead of synergistic effect among their combinations. Additionally, Palafox-Carlos et al examined the radical scavenge capacity (RSA) of polyphenol vanillic acid (A), protocatechuic acid (B), and chlorogenic acid (C) using 2,2-diphenyl-1-picrylhydrazy (DPPH) assay, resulting in RSAs of 5.6%, 17.30%, and 13.56%, respectively

(32). Meanwhile, the combination of the latter with the polyphenol showed 23.7% (B+A) and 20.4% (C+A) inhibition, respectively (32). Not only in those chemical constituents, but the additive mode has also been proven by the antioxidant assays in binary and ternary fruit extract combinations based on FRAP (ferric reducing antioxidant power), DPPH (2,2-diphenyl-1-picryl-hydrazyl-hydrate), and ORAC (oxygen radical absorbance capacity) assays (38). The addition of antioxidant capacities of individual phenolic compounds led to a total antioxidant capacity that exhibited an effect consistent with their summations from individual components, correlating with an additive action mode. Therefore, instead of relying on limited compounds as the principal active compound, it should be more logical to assume that there is probably a large orchestral mechanism among chemical constituents in plant material working on this target, which produces additive interactions (39). Hence, considering gradual effect as the main priority based on the bioassay-guided fractionation model must be reevaluated as the standard for finding pharmaceutical active materials. From an economic point of view, leveraging crude extracts for pharmaceutical dosage forms is more economical than the use of their fractions or purified compounds (40,41).

# The role of aqueous extracts

In term of solvent choice, several investigations have shown that aqueous extracts from certain plant species display more pronounced activity compared to those obtained using organic solvents (42-47). A considerable amount of glycoside derivatives of secondary metabolites can be identified in aqueous extracts (48,49). However, these aqueous extracts mainly consist of primary metabolites, including peptides, carbohydrates, and certain inorganic compounds. In this regard, traditional preparation techniques for herbal medicine include decoction, boiling of dried plants, or expression of fresh material, which may attract predominantly primary instead of solely secondary metabolite groups (50,51). In fact, throughout history, long before the discovery of organic solvents for extraction, it has been widely recognized that traditional preparations involved extracting essential components from fresh herbs parts through methods such as juicing of fresh plant materials (52) or the utilization of plant materials in the form of decoctions or infusions with water (53,54). Therefore, it is plausible that the effectiveness of traditional infusion or decoction preparations arises from the interaction of primary and secondary metabolites, rather than from the effects of secondary metabolites alone.

# Primary and secondary metabolites

Unfortunately, the majority of research papers predominantly have been focusing on the toxic aspects of metals and minerals in herbal products (55-57), leading

to a scarcity of reports on the bioactivity of these minerals and trace elements in phytomedicine. However, the long-term consumption of these materials, particularly those present in herbal teas such as Al, Ba, Fe, Zn, Mn, Mg, K, Na, P, Cu, Sr, and Ca has been fundamentally acknowledged as playing a crucial role in human growth and overall health (58,59). Furthermore, some of these elements are vitally important for various preventive and promotive health states in human body (60,61). It is also widely recognized that minerals, when present in low concentrations, are soluble in water and may coexist with secondary or primary metabolites in traditional infusions or decoctions. Zhang et al reported that a synergistic effect occurs between Zn, Ca, Mn and flavonoids from Epimedium koreanum extract, which stimulates primary osteoblast by enhancing alkaline phosphatase activity (62). The combination of green tea extract containing rich polyphenols with Se, Cu, Mn and some amino acids demonstrated a significant inhibition of the growth of many cancer cell models (63,64). Trace elements such as Zn, Fe, and Cu, which are found in *Morus alba* extract (65), along with Cr and Mg in Origanum grosii leaf extract, are suggested to exhibit a synergistic antihyperglycemic effect when combined with the secondary metabolites present in these extracts (66). Biel et al reported that with varying content of Zn, Fe, Cr, and Mn in Cynara scolymus leaf extract, when interacting with a high polyphenol content and fixed carbohydrates, fats, and proteins, may contribute to the prevention of chronic non-communicable diseases, through protection from oxidative damage mechanism (67). Regarding the role of primary metabolites, particularly polysaccharide, Cho et al reported that a 3:1 combination of glucan polymer from Aureobasidium pullulans and Textoria morbifera in ovariectomized mice exhibit the strongest anti-osteoporotic effect (68). The combination of selenium with natural polyphenols, such as resveratrol (69,70) and chlorogenic acid (71) has been revealed to inhibit metal-induced Aß aggregation involved in the development of Alzheimer's disease. Therefore, both secondary and primary metabolites may exert a potentially significant impact on various pathological conditions, particularly in the form of decoctions and infusions (68,72-74). Therefore, depending on the linked pharmacology class, it is urgent to provide the mineral and trace element profiles in herbal preparations, too. A summary of possible coworking action modes among secondary, primary, complex mixtures, and minerals is presented in Table 1.

# Putative modeling of the pharmacological mechanisms of herbal preparations

Based upon those bioactivity mode overviews, relying on conventional pharmacological concepts, "one active compound toward one target" apparently is inappropriate to be applied to herbal preparations. To bridge the gap,

| Table 1. | Cooperative | action mode | among | secondary | metabolites, | primary | metabolites, | and mi | nerals |
|----------|-------------|-------------|-------|-----------|--------------|---------|--------------|--------|--------|
|----------|-------------|-------------|-------|-----------|--------------|---------|--------------|--------|--------|

| Component involved                                 | Compound groups and bioactivity   | Putative<br>mechanism  | Bioassay<br>type | References |
|--|---|------------------------|------------------|------------|
|  | Berberine and piperine: antibacterial   | Synergist              | In vitro         | (28)       |
| Secondary<br>metabolites                           | Artemisinin and flavonoids: antimalaria   | Synergist              | In vitro         | (29)       |
|  | Saponins and flavonoids: myocardial ischemia  | Synergist              | In vivo          | (75)       |
|  | Prenylated xanthonoids (acetylshikonin, deoxyshikonin, $\beta$ ,<br>$\beta$ -dimethylacrylshikonin, $\beta$ -hydroxyisovalerylshikonin, and trans-anethole):<br>cell regenerating stimulation for wound healing | Additive               | In vitro         | (76)       |
|  | Berberine, hypaconitine and skimmianine: ulcerative colitis   | Complementary          | In vivo          | (77)       |
|  | Tanshinone IIA terpenoid, salvianolic acid B and ginsenoside Rb1 saponin: myocardial ischemia   | Synergist              | In vivo          | (78)       |
|  | Anthocyanidin and rosmarinic acid: anti colitis   | Synergist              | In vivo          | (79)       |
| Secondary metabolite<br>with primary<br>metabolite | Ginsenoside Rb1 and polysaccharides; Diabetic model and antistress  | Synergist              | In vivo          | (80,81)    |
|  | Lignan and polysaccharide; hepatoprotective   | Synergist              | In vivo          | (82)       |
|  | Puerarin and polysaccharide: diabetes type II   | Synergist              | In vivo          | (83)       |
|  | Polysaccharide with arabinogalactan type II side chains of Piper nigrum fruits and its synergistic effect with piperine; antitussive  | Synergist              | In vivo          | (84)       |
|  | Flavonoids and polysaccharides; dementia  | Synergist              | In vivo          | (85)       |
|  | Polysaccharides of Auricularia auricula and tremella and flavonoid; anti-<br>dyslipidemia   | Synergist              | In vivo          | (86)       |
|  | Polyphenols and omega-3 fatty acid; antidepressant  | Synergist              | In vivo          | (87)       |
|  | Procyanidin B3 and eicosapentaenoic acid<br>Polyunsaturated fatty acids; anti-inflammatory  | Synergist              | In vitro         | (88)       |
|  | Among polysaccharides; anti-inflammatory  | Synergist              | In vitro         | (89)       |
| Among primary<br>metabolites                       | Among polysaccharides   | Synergist              | In vitro         | (90)       |
|  | Glucagon peptide' glucose lowering effect   | Synergist              | In vivo          | (91)       |
|  | Sulfated polysaccharides: anti-measles virus  | Synergist              | In vitro         | (92)       |
|  | Secondary metabolites with polysaccharide with fatty acids or peptide with fatty acids  | No report              |                  |            |
| Secondary metabolite mineral/metal ions            | Flavonoids and trace element ion of B, Cu, Co Mn, Zn: general health promotion  | Possible complementary | -                | (93)       |

more extended pharmacological mechanism theories may be proposed. There are four plausible mechanism concepts that could be approached. The first theory is based on the major compound effects present in herbal formulation. This concept follows conventional pharmacology, the pharmacological effects are produced by the binding of the major compound to a specific biomolecule such as an enzyme or receptor. The second idea is synergistic interactions. This is when one or more compounds, through specific actions, boost the activity of an active compound in an herbal preparation, as shown in many reports, including the most recent one by Vidar et al (28). The compound helpers have no effect or relatively do not have a significant effect on a specific target when each metabolite is examined separately. Meanwhile, when they are mixed in a dosage form, the active compound has a significant effect. The third form is additive interaction, as results of coworking of compounds which have similar working target, either enzymes, receptors,

or other biomolecules. When each compound is tested at a certain dose individually against a specific target, the output becomes low because all targets are not occupied. However, it should be noted here that more detailed mechanisms, such as partial agonis, allosteric, and so on, are outside of this review's coverage. On the other hand, when all compounds are formulated in a dosage form, it will result in a maximum effect, which might be due to a mode of action combination such as agonist and allosteric or due to total occupancy of the target.

#### Reintroducing complementary effect

The last plausible coworking form is the complementary effect, as mentioned by Zhang et al (77). This coworking action mode involves multiple compounds with multiple targets, all of which are directed to produce the same main outcome of a certain pathological class improvement. As with the active drugs in hypertension therapy, there are different drug classes, e.g., angiotensin converting enzyme (ACE) inhibitor, angiotensin receptor inhibitor, diuretic, and so on (77,94-96). Those agents have different specific targets, but all those sub-mechanisms are directed to lower blood pressure. Furthermore, because this complementary mode of action consists of proposed complex biological pathways, it has become the current pharmacology network paradigm (97,98). Nevertheless, those interactions must consider the concentration and ratio of the involving compounds.

#### Drawback of current metabolomic techniques

Over the last decade metabolomic techniques have emerged in response to the demand for a pattern-oriented approach that bridges the gap between holistic and comprehensive perspectives (99). However, a significant portion of reports has focused on small molecule groups and short-chain primary metabolites such as short sugars, amino acids, and short fatty acids (100,101), whilst their output data often exhibit limited direct correlations with bioactivity data. On the other hand, macromolecules (polysaccharides, peptides, and fatty acids), conjugates, and minerals in the herbal preparations might have bioactivity contributions that mainstream analytical methods have overlooked (102-105). Considering the secondary metabolite platform, therefore, a point to argue here is that there is some missing information from whole connectivity between the total metabolites and biological activities (106,107). Despite still being at the level of preclinical studies, the number of synergistic interaction reports between primary metabolite and other chemical constituent groups are relatively supportive of their existence (108).

### Contribution of non-secondary metabolites

Shi et al (86) reported that polysaccharides of edible fungal Auricularia and Tremellan and flavonoid of Crataegus sp. demonstrated a synergistic effect as anti-dyslipidemia in vivo. Yan et al (82) reported that the combination of lignan from Schisandrae chinensis fruits and polysaccharide from Astragalus sp. showed a synergistic interaction as a hepatoprotective effect on a rat model. The combination of polysaccharide with arabinogalactan type II side chains and piperine was reported to have a synergistic effect as a cough suppressor (84). In vivo tests of the combination of puerarin and polysaccharide from Pumpkin (109) and polysaccharides and ginsenoside Rb1 from Panax ginseng (80) exhibited some synergistic effects as antidiabetic type 2. Fang et al (85) reported that ginkgo flavonoids and Coriolus versicolor polysaccharides showed synergistic effects as antidementia agents in a rat model. The coworking effect between secondary metabolites and fatty acids was demonstrated by the combinations of polyphenols and omega-3 fatty acids as synergistic antidepressants (87) and procyanidin B3 and eicosapentaenoic acid as antiinflammatory agents (88). Meanwhile, the cooperative

action mode between polysaccharides and peptides was reported on glucagon peptide as glucose lowering effect of Ilex paraguariensis (91) and conjugated polysaccharides polypeptide, hypoglycemic, hypolipidemicwith antiatherogenic, anticoagulant, and antithrombogenic agents (110). The report of possible cooperative action mode between secondary metabolite and minerals has been revealed on flavonoids and trace element ion of Cu, Co, Mn, or Zn in the composition of tea infusion (93). However, the current reports on potential coworking action modes predominantly are still at in vivo level. It is understandable since the "wet laboratory work" of natural product studies is mainly directed toward bioassay guided isolation for lead compounds. As a consequence, research reports on the macromolecules and mineral activity and their interaction toward secondary metabolites as well as their specific mechanism in herbal pharmacology are found quite rare. Nevertheless, from those ample of reported data, the contributions of polysaccharides, peptides, fatty acids or their conjugates, and minerals toward functional bioactivities to the crude drugs, in particular in aqueous and polar extracts, cannot be ignored. Therefore, once a defined pharmacology class of crude drugs has been established, the standardization requirements must be comprehensively fulfilled for all chemical groups. It is likely an unfair scenario if the standardization of herbal preparations sticks to relying on one type of secondary metabolite group or certain chemical markers as the specific standardization purpose. To address the gap, a comprehensive physical-chemical standardization, including the efforts to accommodate the classical quantification of polysaccharides, peptides, and mineral are indispensable with the initial guide of the biological activities (Figure 1).

### Pondering classic herbal standardizations

Based upon the simplified pharmacological approaches, it implies that the current extract standardization might not adequately represent all contributing chemical constituents, particularly in terms of the terminology of specific parameters (111,112). Therefore, a more farreaching paradigm should be considered. Once biologically active samples have been confirmed, the appropriate chemico-physical profiling methods of all metabolite types and all possible contributing minerals should also be conducted. For conventional extract batch uniformity tests, some investigators use certain chemical markers from secondary metabolite groups, which are called as active markers with HPTLC or HPLC quantification methods (113-115). Nevertheless, the efficacy of active markers is insufficient to show how all chemicals in certain plant species or even multiple components work together. This concept is only relevant to the active major compounds that have been identified so far. For this reason, the active markers very often fall into the category



Figure 1. Flow chart of combination chemical finger printing and classical quantification of all chemical constituent groups.

of analytical markers without any correlation to quality, in particular efficacy (116). It is then clear that patternoriented approaches are more appropriate methodologies for possible synergistic, additive, and complementary interactions of compounds in herbal preparations. Current hyphenated instruments, in particular LC-MS and LC-NMR, in combination with chemometric platforms become choices as possibilities to provide "holistic or comprehensive views" in some senses (117,118). However, rather than a single system for all molecule groups, LC-MS instrumentations employ different solvents, stationary phases, and detection systems dependent upon the metabolite group in terms of molecular weight to accommodate a wide range of metabolites, including carbohydrates, peptides, fatty acids, and their conjugates. Therefore, for one sample, one might use several LC-MS types (79,119). Even though NMR spectrometry can cover all molecule groups with one spectrum, it is hard to imagine how many metabolite signals will overlap (120,121), and the sensitivity parameter and solvent choice solubility strength (122) become problems in the NMR method. Nonetheless, regarding the cost, in particular, LC-MS and NMR-based methods become issues for third-world countries. Considering these reasons, the scarcity, the high cost of the reference substances, and advanced instrumentation availability have been the three major bottlenecks in the implementation of multicomponent routine quality control and pattern-oriented

approach of herbal standardization. Thus, the extended standardization methods which align effectively with traditional practices, like the solubility of the total extract and ash in water, semi-polar solvents, and nonpolar solvents (Figure 1), could be reintroduced as the element of complete standardization approaches.

The traditional standardization methods based on colorimetry or gravimetry methods of total quantification of each metabolite group, such as flavonoid, polyphenol, alkaloid, tannin, coumarin, glucosinolate, protein, carbohydrate, fatty acid, and peptide groups, therefore, are still useful to fix the lack of thoroughness. Until recently, the vast majority of metal analysis in herbal medicines has been directed towards investigating their toxicity rather than recognizing their potential beneficial and fundamental contribution to human health. Further, assessing the total quantity of a specific mineral in an extract based on inductively coupled plasma-mass spectrometry (ICP-MS) could be applied since the interaction among any type of chemical constituent could be considered (123,124). ICP-MS is the current method of choice to quantify certain minerals or targeted mineral groups in very specific and sensitive quantification with a lower detection limit down to part per trillion (ppt) with efficient performance. Meanwhile, relatively uncovered matters by advanced analytical instruments in extract can be quantified by gravimetry methods with solubility division successively in water and ethanol prior

to the constant weight quantification. Ash characters representing whole acidic and basic minerals are dissolved in corresponding solvents before determining each constant weight according to USP (125). The chart of chemical group analysis and classical quantification of all chemical constituent groups is summarized in Table 2.

# Periodic standardization and metabolite evolution

In addition to good agricultural practice implementations, considering the variability of regional resources and evolution's impact, official standards such as herbal pharmacopeia and monograph should be routinely evaluated every five to eight years. Several factors may trigger the secondary metabolite change due to hybridization (147), variability of herbivore-plant interaction (148) and insect resistance, and other environmental changes such as drought stress, salinity, higher-lower temperature, and soil microbial composition (149,150). These conditions will impact chemo-diversity on both metabolite composition and accumulation (151,152).

# Conclusion

The bioassay guided fractionation program was only able to result in 0.01% clinically used drugs; besides, the purified matters have exhibited lower efficacy than that of the parent extracts. Thus, leveraging crude extracts will be more beneficial as the main raw material for health supporting products. Since the evidence based on herbal medicine is relatively lacking in data, a plethora of herbal medicine studies have been focused on secondary

metabolites or macromolecules, which might not be so relevant to the traditional context, which is apparently based on complex constituents. As a consequence, once the bioactivity of an herbal extract is defined, conducting all aspects of chemical constituent groups is mandatory. Too much focus on the secondary metabolite group is apparently irrelevant to the basic pharmacological mechanism approach, which could result from multicomponent interactions in the numerous biomolecular targets. Therefore, the quantification of primary metabolite groups such as polysaccharides, peptides, and fatty acids, as well as mineral content, will complete the data. Traditional methods such as gravimetry for constant weight determination and solubility in acidic or basic solvents of extract and ash are relevant to justify the basic character of material bulk and these methods have been unchangeable until now. Since ecological and climate changes could interfere with the chemical constituent in plant source, the periodic standardization may be conducted to provide official up dating on the official guides.

#### Acknowledgement

The authors are grateful to Pura Pharm Co. Ltd. for funding the project under the Association for Promoting Sustainable Use of Medicinal Resources (APSUMR) 2021.

#### Authors' contribution

**Conceptualization:** Azis Saifudin, Muh. Akbar Bahar, Yasuhiro Tezuka, and Ken Tanaka

Data curation: Azis Saifudin, Muh. Akbar Bahar, Yasuhiro Tezuka, and Ken Tanaka.

 Table 2. Chemical constituent groups and quantification methods

| Constituent item                  | Method  | References   |
|-----------------------------------|---|--------------|
| Total polysaccharide              | Phenol-sulfuric acid method, Colorimetry, Near-infrared hyperspectral imaging       | (126-129)    |
| Total fatty acids                 | One step extraction-esterification  | (130,131)    |
| Total peptides                    | Kjeldahl method, Spectrometry UV, Extraction with TCA-acetone                       | (132-134)    |
| Specific mineral                  | ICP (inductively coupled plasma) Spectroscopy, AAS (atomic absorption spectrometry) | (135,136)    |
| Total water-soluble extract       | Gravimetry  | (125)        |
| Total alcohol-soluble extract     | Gravimetry  | (125)        |
| Ash constant weight               | Ignition method-gravimetry infrared spectrometry                                    | (137)        |
| Total ash soluble water           | Fractionation-gravimetry  | (125)        |
| Total ash soluble in acidic water | Fractionation-gravimetry  | (125)        |
| Total ash soluble in basic water  | Fractionation-gravimetry  | (125)        |
| Total polyphenol                  | Spectrophotometry   | (138)        |
| Total flavonoid                   | Spectrophotometry   | (93,118,139) |
| Total coumarin                    | Spectrophotometry   | (140)        |
| Total steroid                     | Spectrophotometry   | (141)        |
| Total alkaloid                    | Gravimetry  | (142-144)    |
| Total glucosinolates              | High performance liquid chromatography  | (145,146)    |

#### Saifudin et al

Formal analysis: Muh. Akbar Bahar, Yasuhiro Tezuka, and Ken Tanaka.

**Funding acquisition:** Azis Saifudin and Muh. Haqqi Hidayatullah. **Investigation:** Azis Saifudin, Muh. Akbar Bahar, Yasuhiro Tezuka, and Ken Tanaka

Methodology: Azis Saifudin, Muh. Akbar Bahar, Yasuhiro Tezuka, and Ken Tanaka.

Project administration: Muh. Haqqi Hidayatullah.

Resources: Hisayoshi Norimoto, Yasuhiro Tezuka, and Ken Tanaka.

**Software:** Muh. Akbar Bahar, Yasuhiro Tezuka, and Ken Tanaka. **Supervision:** Hisayoshi Norimoto, Yasuhiro Tezuka and Ken Tanaka.

Validation: Muh. Akbar Bahar, Yasuhiro Tezuka, and Ken Tanaka. Visualization: Muh. Haqqi Hidayatullah, and Muh. Akbar Bahar. Writing-original draft: Azis Saifudin and Muh. Akbar Bahar.

Writing-review and editing: Muh. Akbar Bahar, Hisayoshi Norimoto, Yasuhiro Tezuka, and Ken Tanaka.

#### **Conflict of interests**

The authors declare no conflict of interest.

#### **Ethical considerations**

The ethical clearance (number EC. No. 5116.2023) was approved by Health Research Committee of Universitas Muhammadiyah Surakarta according to the Helsinki Declaration 1975, Council for International Organizations of Medical Sciences (CIOMS), and World Health Organization (WHO) 2016.

#### **Funding/Support**

The study was funded by the Association for Promoting Sustainable Use of Medicinal Resources (APSUMR) 2021, a collaboration scheme among Universitas Muhammadiyah Surakarta, Ritsumeikan University, Hokuriku University, and PuraPharm Co. Ltd.

#### References

- Salim AA, Chin YW, Kinghorn AD. Drug discovery from plants. Bioact Mol Med Plants. 2008;1–24. doi.org: 10.1007/978-3-540-74603-4.
- Demain AL. Importance of microbial natural products and the need to revitalize their discovery. J Ind Microbiol Biotechnol. 2014;41(2):185–201. doi.org: 10.1007/s10295-013-1325-z
- Newman DJ, Cragg GM. Natural products as sources of new drugs over the last 25 years. J Nat Prod. 2007;70(3):461–77. doi:10.1021/acs.jnatprod.5b01055
- Cragg GM, Newman DJ. Natural products: a continuing source of novel drug leads. Biochim Biophys Acta BBA-Gen Subj. 2013;1830(6):3670–95. doi: 10.1016/j. bbagen.2013.02.008
- Newman DJ, Cragg GM. Natural products as sources of new drugs over the nearly four decades from 01/1981 to 09/2019. J Nat Prod. 2020;83(3):770–803. doi: 10.1021/acs. jnatprod.9b01285
- Li Y, Shen Y, Yao C liang, Guo D an. Quality assessment of herbal medicines based on chemical fingerprints combined with chemometrics approach: A review. J Pharm Biomed Anal. 2020;185:113215. doi: 10.1016/j.jpba.2020.113215
- 7. Sahoo N, Manchikanti P, Dey S. Herbal drugs: standards

and regulation. Fitoterapia. 2010;81(6):462-71. doi: 10.1016/j.fitote.2010.02.001

- Raidron C, Jordaan A, Seldon R, Warner DF, de Kock C, Taylor D, et al. Antiplasmodial and antimycobacterial activities of crude and lead-like enhanced extracts from Namibian medicinal plants. J Ethnopharmacol. 2022;295:115389. doi: 10.1016/j.jep.2022.115389
- Dai Y, Chen SR, Chai L, Zhao J, Wang Y, Wang Y. Overview of pharmacological activities of *Andrographis* paniculata and its major compound andrographolide. Crit Rev Food Sci Nutr. 2019;59(sup1):S17-29. doi: 10.1080/10408398.2018.1501657
- Kojima-Yuasa A, Matsui-Yuasa I. Pharmacological Effects of 1'-Acetoxychavicol Acetate, a Major Constituent in the Rhizomes of *Alpinia galanga* and *Alpinia conchigera*. J Med Food. 2020;23(5):465-75. doi: 10.1089/jmf.2019.4490
- Kamatou GP, Viljoen AM. Linalool-A review of a biologically active compound of commercial importance. Nat Prod Commun. 2008;3(7):1934578X0800300727. doi: 10.1177/1934578X0800300727
- Adnan M, Patel M, Reddy MN, Alshammari E. Formulation, evaluation and bioactive potential of *Xylaria primorskensis* terpenoid nanoparticles from its major compound xylaranic acid. Sci Rep. 2018;8(1):1740. doi: 10.1038/s41598-018-20237-z
- Barnes J, Anderson LA, Phillipson JD. St John's wort (*Hypericum perforatum* L.): a review of its chemistry, pharmacology and clinical properties. J Pharm Pharmacol. 2001;53(5):583-600. doi: 10.1211/0022357011775910
- Weller MG. A unifying review of bioassay-guided fractionation, effect-directed analysis and related techniques. Sensors. 2012;12(7):9181-209. doi: 10.3390/ s120709181
- Win NN, Awale S, Esumi H, Tezuka Y, Kadota S. Bioactive secondary metabolites from *Boesenbergia pandurata* of Myanmar and their preferential cytotoxicity against human pancreatic cancer PANC-1 cell line in nutrient-deprived medium. J Nat Prod. 2007;70(10):1582-7. doi: 10.1021/ np070286m
- Win NN, Awale S, Esumi H, Tezuka Y, Kadota S. Novel anticancer agents, kayeassamins C- I from the flower of *Kayea assamica* of Myanmar. Bioorg Med Chem. 2008;16(18):8653-60. doi: 10.1016/j.bmc.2008.07.091
- Poongunran J, Perera HKI, Jayasinghe L, Fernando IT, Sivakanesan R, Araya H, et al. Bioassay-guided fractionation and identification of α-amylase inhibitors from *Syzygium cumini* leaves. Pharm Biol. 2017;55(1):206-11. doi: 10.1080/13880209.2016.1257031
- Saifudin A, Tanaka K, Kadota S, Tezuka Y. Protein tyrosine phosphatase 1B (PTP1B)-inhibiting constituents from the leaves of *Syzygium polyanthum*. Planta Med. 2012;78(12):1378-81. doi: 10.1055/s-0032-1315000
- Ding W, Zhang X, Yin X, Zhang Q, Wang Y, Guo C, et al. Ganoderma lucidum aqueous extract inducing PHGPx to inhibite membrane lipid hydroperoxides and regulate oxidative stress based on single-cell animal transcriptome. Sci Rep. 2022;12(1):3139. doi: 10.1038/s41598-022-06985-z
- Gill H, Sykes EM, Kumar A, Sorensen JL. Isolation of bioactive metabolites from soil derived fungus-Aspergillus fumigatus. Microorganisms. 2023;11(3):590. doi: 10.3390/

microorganisms11030590

- 21. Castañeda-Ramírez GS, de Jesús Torres-Acosta JF, Sandoval-Castro CA, Borges-Argáez R, Cáceres-Farfán M, Mancilla-Montelongo G, et al. Bio-guided fractionation to identify *Senegalia gaumeri* leaf extract compounds with anthelmintic activity against *Haemonchus contortus* eggs and larvae. Vet Parasitol. 2019;270:13-9. doi: 10.1016/j. vetpar.2019.05.001
- 22. Le DD, Yu S, Dang T, Lee M. Molecular Networking and Bioassay-Guided Preparation and Separation of Active Extract and Constituents from *Vicia tenuifolia* Roth. Antioxidants. 2023;12(10):1876. doi: 10.3390/ antiox12101876
- 23. Yang J, Hu DB, Xia MY, Luo JF, Li XY, Wang YH. Bioassayguided isolation of cytotoxic constituents from the flowers of *Aquilaria sinensis*. Nat Prod Bioprospecting. 2022;12(1):11. doi: 10.1007/s13659-022-00334-3
- Gao Y, He C, Bi W, Wu G, Altman E. Bioassay guided fractionation identified hederagenin as a major cytotoxic agent from *Cyclocarya paliurus* leaves. Planta Med. 2016;82(1-2):171-9. doi: 10.1055/s-0035-1557900
- Yang Y, Cheng X, Liu W, Chou G, Wang Z, Wang C. Potent AChE and BChE inhibitors isolated from seeds of *Peganum harmala* Linn by a bioassay-guided fractionation. J Ethnopharmacol. 2015;168:279-86. doi: 10.1016/j. jep.2015.03.070
- 26. Ohizumi Y, Kawada M, Kamada M, Nakajima A, Kajima K, Uozumi N, et al. Isolation of adenosine and cordysinin B from *Anredera cordifolia* that stimulates CRE-mediated transcription in PC12 cells. Planta Medica Int Open. 2021;8(01):e19-24. doi: 10.1055/a-1395-6510
- Esmaeili F, Farhadpour M, Abbas-Mohammadi M, Alilou M, Morshedi D, Ebrahimie E, et al. Appraisals on the anticancer properties of Mentha species using bioassays and docking studies. Ind Crops Prod. 2023;203:117128. doi: 10.1016/j.indcrop.2023.117128
- Vidar WS, Baumeister TU, Caesar LK, Kellogg JJ, Todd DA, Linington RG, et al. Interaction Metabolomics to Discover Synergists in Natural Product Mixtures. J Nat Prod. 2023;86(4):655-71. doi: 10.1021/acs.jnatprod.2c00518
- 29. Bilia AR, Sannella AR, Vincieri FF, Messori L, Casini A, Gabbiani C, et al. Antiplasmodial effects of a few selected natural flavonoids and their modulation of artemisinin activity. Nat Prod Commun. 2008;3(12):1934578X0800301212. doi: 10.1177/1934578X0800301212
- Elfawal MA, Towler MJ, Reich NG, Weathers PJ, Rich SM. Dried whole-plant Artemisia annua slows evolution of malaria drug resistance and overcomes resistance to artemisinin. Proc Natl Acad Sci. 2015;112(3):821-6. doi: 10.1073/pnas.1413127112
- Cai TY, Zhang YR, Ji JB, Xing J. Investigation of the component in Artemisia annua L. leading to enhanced antiplasmodial potency of artemisinin via regulation of its metabolism. J Ethnopharmacol. 2017;207:86-91. doi: 10.1016/j.jep.2017.06.025
- 32. Palafox-Carlos H, Gil-Chávez J, Sotelo-Mundo RR, Namiesnik J, Gorinstein S, González-Aguilar GA. Antioxidant interactions between major phenolic compounds found in 'Ataulfo' mango pulp: chlorogenic,

gallic, protocatechuic and vanillic acids. Molecules. 2012;17(11):12657-64. doi: 10.3390/molecules171112657

- 33. Skroza D, Mekinić IG, Svilović S, Šimat V, Katalinić V. Investigation of the potential synergistic effect of resveratrol with other phenolic compounds: A case of binary phenolic mixtures. J Food Compos Anal. 2015;38:13-8. doi: 10.1016/j.jfca.2014.06.013
- Olszowy-Tomczyk M. Synergistic, antagonistic and additive antioxidant effects in the binary mixtures. Phytochem Rev. 2020;19:63-103. doi: 10.1007/s11101-019-09658-4
- Payne RA. The epidemiology of polypharmacy. Clin Med. 2016;16(5):465. doi: 10.7861/clinmedicine.16-5-465
- Masnoon N, Shakib S, Kalisch-Ellett L, Caughey GE. What is polypharmacy? A systematic review of definitions. BMC Geriatr. 2017;17:1-10. doi: 10.1186/s12877-017-0621-2
- Heo HJ, Kim YJ, Chung D, Kim DO. Antioxidant capacities of individual and combined phenolics in a model system. Food Chem. 2007;104(1):87-92. doi: 10.1016/j. foodchem.2006.11.002
- Wang S, Meckling KA, Marcone MF, Kakuda Y, Tsao R. Synergistic, additive, and antagonistic effects of food mixtures on total antioxidant capacities. J Agric Food Chem. 2011;59(3):960-8. doi: 10.1021/jf1040977
- Awouafack MD, McGaw LJ, Gottfried S, Mbouangouere R, Tane P, Spiteller M, et al. Antimicrobial activity and cytotoxicity of the ethanol extract, fractions and eight compounds isolated from *Eriosema robustum* (Fabaceae). BMC Complement Altern Med. 2013;13:1–9. doi: 10.1186/1472-6882-13-289
- Ravber M, Knez Ž, Škerget M. Isolation of phenolic compounds from larch wood waste using pressurized hot water: extraction, analysis and economic evaluation. Cellulose. 2015;22:3359–75. doi: 10.1007/s10570-015-0719-7
- 41. Selvamuthukumaran M, Shi J. Recent advances in extraction of antioxidants from plant by-products processing industries. Food Quality and Safety. 2017;1(1):61-81. doi: 10.1093/fqs/fyx004
- 42. Phillips OA, Mathew KT, Oriowo MA. Antihypertensive and vasodilator effects of methanolic and aqueous extracts of *Tribulus terrestris* in rats. J Ethnopharmacol. 2006;104(3):351-5. doi: 10.1016/j.jep.2005.09.027
- Ismail H, Lemriss S, Aoun ZB, Mhadhebi L, Dellai A, Kacem Y, et al. Antifungal activity of aqueous and methanolic extracts from the Mediterranean sea cucumber, *Holothuria polii*. J Mycol Médicale. 2008;18(1):23-6. doi: 10.1016/j. mycmed.2008.01.002
- Sudha G, Vadivukkarasi S, Shree RBI, Lakshmanan P. Antioxidant activity of various extracts from an edible mushroom *Pleurotus eous*. Food Sci Biotechnol. 2012;21:661-8. doi: 10.1007/s10068-012-0086-1
- 45. Caboni P, Saba M, Tocco G, Casu L, Murgia A, Maxia A, et al. Nematicidal activity of mint aqueous extracts against the root-knot nematode *Meloidogyne incognita*. J Agric Food Chem. 2013;61(41):9784-8. doi: 10.1021/jf403684h
- 46. Pawlowicz K, Paczkowska-Walendowska M, Osma lek T, Cielecka-Piontek J. Towards the preparation of a hydrogel from lyophilisates of the *Aloe arborescens* aqueous extract. Pharmaceutics. 2022;14(7):1489. doi: 10.3390/ pharmaceutics14071489

- Cédric Y, Nadia NAC, Nfufu S, Azizi MA, Sandra TNJ, Payne VK, et al. Nematocidal activity of ethanol and aqueous extracts of *Persea americana* seeds against Heligmosomoides polygyrus using the worm microtracker method. J Parasitol Res. 2023;2023:1-6. doi: 10.1155/2023/9545565
- Wu L, Georgiev MI, Cao H, Nahar L, El-Seedi HR, Sarker SD, et al. Therapeutic potential of phenylethanoid glycosides: A systematic review. Med Res Rev. 2020;40(6):2605-49. doi: 10.1002/med.21717
- Bartnik M, Facey P. Glycosides. In: Pharmacognosy. Elsevier; 2024. p. 103-65. doi: 10.1016/B978-0-443-18657-8.00001-3
- 50. Nagai T, Inoue R. Preparation and the functional properties of water extract and alkaline extract of royal jelly. Food Chem. 2004;84(2):181-6. doi: 10.1016/s0308-8146(03)00198-5
- 51. Liu N, Yang M, Huang W, Wang Y, Yang M, Wang Y, et al. Composition, antioxidant activities and hepatoprotective effects of the water extract of *Ziziphus jujuba* cv. Jinsixiaozao. Rsc Adv. 2017;7(11):6511–22. doi: 10.1039/c6ra27516h
- Francia S, Stobart A. Critical approaches to the history of Western herbal medicine: from classical antiquity to the early modern period. Bloomsbury Academic; 2014. p. 304. doi: 10.5040/9781474210577
- Guimarães R, Barros L, Carvalho AM, Ferreira IC. Infusions and decoctions of mixed herbs used in folk medicine: synergism in antioxidant potential. Phytother Res. 2011;25(8):1209-14. doi: 10.1002/ptr.3366
- 54. Liu WJ. Traditional herbal medicine research methods: identification, analysis, bioassay, and pharmaceutical and clinical studies. John Wiley & Sons; 2011. p. 81-138. doi: 10.1002/9780470921340
- 55. Saper RB, Kales SN, Paquin J, Burns MJ, Eisenberg DM, Davis RB, et al. Heavy metal content of ayurvedic herbal medicine products. JAMA. 2004;292(23):2868–73. doi: 10.1001/jama.292.23.2868
- Luo L, Wang B, Jiang J, Fitzgerald M, Huang Q, Yu Z, et al. Heavy metal contaminations in herbal medicines: Determination, comprehensive risk assessments, and solutions. Front Pharmacol. 2021;11:595335. doi: 10.3389/ fphar.2020.595335
- 57. Yang CM, Chien MY, Chao PC, Huang CM, Chen CH. Investigation of toxic heavy metals content and estimation of potential health risks in Chinese herbal medicine. J Hazard Mater. 2021;412:125142. doi: 10.1016/j. jhazmat.2021.125142
- Nookabkaew S, Rangkadilok N, Satayavivad J. Determination of trace elements in herbal tea products and their infusions consumed in Thailand. J Agric Food Chem. 2006;54(18):6939–44. doi: 10.1021/jf060571w
- Tokalıoğlu Ş. Determination of trace elements in commonly consumed medicinal herbs by ICP-MS and multivariate analysis. Food Chem. 2012;134(4):2504–8. doi: 10.1016/j. foodchem.2012.04.093
- Özcan MM, Ünver A, Uçar T, Arslan D. Mineral content of some herbs and herbal teas by infusion and decoction. Food Chem. 2008;106(3):1120-7. doi: 10.1016/j. foodchem.2007.07.042
- 61. Pytlakowska K, Kita A, Janoska P, Po\lowniak M, Kozik V. Multi-element analysis of mineral and trace elements

in medicinal herbs and their infusions. Food Chem. 2012;135(2):494-501. doi: 10.1016/j.foodchem.2012.05.002

- Zhang D, Cheng Y, Zhang J, Wang X, Wang N, Chen Y, et al. Synergistic effect of trace elements and flavonoids from *Epimedium koreanum* Nakai on primary osteoblasts. Chin Sci Bull. 2008;53(3):347-56. doi: 10.1007/s11434-007-0485-5
- Roomi M, Monterrey J, Kalinovsky T, Rath M, Niedzwiecki A. Comparative effects of EGCG, green tea and a nutrient mixture on the patterns of MMP-2 and MMP-9 expression in cancer cell lines. Oncol Rep. 2010;24(3):747-57. doi: 10.3892/or\_00000917
- Niedzwiecki A, Roomi MW, Kalinovsky T, Rath M. Anticancer efficacy of polyphenols and their combinations. Nutrients. 2016;8(9):552. doi: 10.3390/nu8090552
- 65. Król E, Jeszka-Skowron M, Krejpcio Z, Flaczyk E, Wójciak RW. The effects of supplementary mulberry leaf (*Morus alba*) extracts on the trace element status (Fe, Zn and Cu) in relation to diabetes management and antioxidant indices in diabetic rats. Biol Trace Elem Res. 2016;174:158-65. doi: 10.1007/s12011-016-0696-1
- 66. El Hassouni H, Bouhrim M, El Hajji R, Bnouham M, Ziyyat A, Romane A. Characterization of an endemic plant *Origanum grosii* from Morocco: trace element concentration and Antihyperglycemic activities. J Chem. 2021;2021:1-10. doi: 10.1155/2021/8840998
- Biel W, Witkowicz R, Piątkowska E, Podsiadlo C. Proximate composition, minerals and antioxidant activity of artichoke leaf extracts. Biol Trace Elem Res. 2020;194:589-95. doi: 10.1007/s12011-019-01806-3
- Cho CS, Jeong HS, Kim IY, Jung GW, Ku BH, Park DC, et al. Anti-osteoporotic effects of mixed compositions of extracellular polymers isolated from *Aureobasidium pullulans* and *Textoria morbifera* in ovariectomized mice. BMC Complement Altern Med. 2018;18:1-15. doi: 10.1186/ s12906-018-2362-y
- 69. Vicente-Zurdo D, Romero-Sánchez I, Rosales-Conrado N, León-González ME, Madrid Y. Ability of selenium species to inhibit metal-induced A $\beta$  aggregation involved in the development of Alzheimer's disease. Anal Bioanal Chem. 2020;412:6485-97. doi: 10.1007/s00216-020-02644-2
- 70. Yang L, Wang Y, Zheng G, Li Z, Mei J. Resveratrol-loaded selenium/chitosan nano-flowers alleviate glucolipid metabolism disorder-associated cognitive impairment in Alzheimer's disease. Int J Biol Macromol. 2023;239:124316. doi: 10.1016/j.ijbiomac.2023.124316
- 71. Li Z, Zheng G, Wang N, Liang H, Li C, Wang Y, et al. A Flower-like Brain Targeted Selenium Nanocluster Lowers the Chlorogenic Acid Dose for Ameliorating Cognitive Impairment in APP/PS1 Mice. J Agric Food Chem. 2023;71(6):2883-97. doi: 10.1021/acs.jafc.2c06809
- Suliburska J, Kaczmarek K. Herbal infusions as a source of calcium, magnesium, iron, zinc and copper in human nutrition. Int J Food Sci Nutr. 2012;63(2):194-8. doi: 10.3109/09637486.2011.617359
- Samolińska W, Kiczorowska B, Kwiecień M, Rusinek-Prystupa E. Determination of minerals in herbal infusions promoting weight loss. Biol Trace Elem Res. 2017;175:495-502. doi: 10.1007/s12011-016-0790-4
- 74. Zhang ZF, Yang JL, Jiang HC, Lai Z, Wu F, Liu ZX. Updated

association of tea consumption and bone mineral density: A meta-analysis. Medicine (Baltimore). 2017;96(12):e6437. doi: 10.1097/MD.0000000006437

- 75. Parvez MK, Al-Dosari MS, Arbab AH, Al-Rehaily AJ, Abdelwahid MA. Bioassay-guided isolation of anti-hepatitis B virus flavonoid myricetin-3-O-rhamnoside along with quercetin from *Guiera senegalensis* leaves. Saudi Pharm J. 2020;28(5):550-9. doi: 10.1016/j.jsps.2020.03.006
- 76. Safavi F, Moridi Farimani M, Golalipour M, Bayat H. In vitro wound healing potential of cyclohexane extract of *Onosma dichroantha* Boiss. based on bioassay-guided fractionation. Sci Rep. 2023;13(1):5018. doi: 10.1038/s41598-023-31855-7
- 77. Zhang M, Long Y, Sun Y, Wang Y, Li Q, Wu H, et al. Evidence for the complementary and synergistic effects of the threealkaloid combination regimen containing berberine, hypaconitine and skimmianine on the ulcerative colitis rats induced by trinitrobenzene-sulfonic acid. Eur J Pharmacol. 2011;651(1–3):187–96. doi: 10.1016/j.ejphar.2010.10.030
- Lu Y, Liu X, Liang X, Xiang L, Zhang W. Metabolomic strategy to study therapeutic and synergistic effects of tanshinone IIA, salvianolic acid B and ginsenoside Rb1 in myocardial ischemia rats. J Ethnopharmacol. 2011;134(1):45–9. doi: 10.1016/j.jep.2010.11.048
- 79. Zhao L, Zhang Y, Liu G, Hao S, Wang C, Wang Y. Black rice anthocyanin-rich extract and rosmarinic acid, alone and in combination, protect against DSS-induced colitis in mice. Food Funct. 2018;9(5):2796–808. doi: 10.1039/ C7FO01490B
- Li J, Li R, Li N, Zheng F, Dai Y, Ge Y, et al. Mechanism of antidiabetic and synergistic effects of ginseng polysaccharide and ginsenoside Rb1 on diabetic rat model. J Pharm Biomed Anal. 2018;158:451–60. doi: 10.1016/j. jpba.2018.06.024
- Shen H, Gao XJ, Li T, Jing WH, Han BL, Jia YM, et al. Ginseng polysaccharides enhanced ginsenoside Rb1 and microbial metabolites exposure through enhancing intestinal absorption and affecting gut microbial metabolism. J Ethnopharmacol. 2018;216:47–56. doi: 10.1016/j.jep.2018.01.021
- 82. Yan F, Zhang QY, Jiao L, Han T, Zhang H, Qin LP, et al. Synergistic hepatoprotective effect of *Schisandrae lignans* with *Astragalus polysaccharides* on chronic liver injury in rats. Phytomedicine. 2009;16(9):805–13. doi: 10.1016/j. phymed.2009.02.004
- Chen X, Qian L, Wang B, Zhang Z, Liu H, Zhang Y, et al. Synergistic hypoglycemic effects of pumpkin polysaccharides and puerarin on type II diabetes mellitus mice. Molecules. 2019;24(5):955. doi: 10.3390/ molecules24050955
- 84. Khawas S, Nosáľová G, Majee SK, Ghosh K, Raja W, Sivová V, et al. In vivo cough suppressive activity of pectic polysaccharide with arabinogalactan type II side chains of *Piper nigrum* fruits and its synergistic effect with piperine. Int J Biol Macromol. 2017;99:335–42. doi: 10.1016/j. ijbiomac.2017.02.093
- 85. Fang X, Jiang Y, Ji H, Zhao L, Xiao W, Wang Z, et al. The synergistic beneficial effects of ginkgo flavonoid and coriolus versicolor polysaccharide for memory improvements in a mouse model of dementia. Evid Based Complement Altern Med. 2015;128394(9). doi: 10.1155/2015/128394

- 86. Shi K, Zhou T, Yuan Y fei, Li D dan, Gong B bin, Gao S, et al. Synergistic mediating effect of edible fungal polysaccharides (Auricularia and Tremellan) and Crataegus flavonoids in hyperlipidemic rats. Food Sci Nutr. 2023;11:4812-4828. doi: 10.1002/fsn3.3459
- Naveen S, Siddalingaswamy M, Singsit D, Khanum F. Antidepressive effect of polyphenols and omega-3 fatty acid from pomegranate peel and flax seed in mice exposed to chronic mild stress. Psychiatry Clin Neurosci. 2013;67(7):501–8. doi: 10.1111/pcn.12100
- Pallares V, Calay D, Cedó L, Castell-Auví A, Raes M, Pinent M, et al. Additive, antagonistic, and synergistic effects of procyanidins and polyunsaturated fatty acids over inflammation in RAW 264.7 macrophages activated by lipopolysaccharide. Nutrition. 2012;28(4):447–57. doi: 10.1016/j.nut.2011.07.027
- Jen CI, Su CH, Lu MK, Lai MN, Ng LT. Synergistic anti-inflammatory effects of different polysaccharide components from *Xylaria nigripes*. J Food Biochem. 2021;45(4):e13694. doi: 10.1111/jfbc.13694
- 90. Deng Y, Xie J, Luo Z, Li SP, Zhao J. Synergistic immunomodulatory effect of complex polysaccharides from seven herbs and their major active fractions. Int J Biol Macromol. 2020;165:530–41. doi: 10.1016/j. ijbiomac.2020.09.199
- 91. Hussein GME, Matsuda H, Nakamura S, Hamao M, Akiyama T, Tamura K, et al. Mate tea (*Ilex paraguariensis*) promotes satiety and body weight lowering in mice: involvement of glucagon-like peptide-1. Biol Pharm Bull. 2011;34(12):1849–55. doi: 10.1248/bpb.34.1849
- 92. Morán-Santibañez K, Cruz-Suárez LE, Ricque-Marie D, Robledo D, Freile-Pelegrín Y, Peña-Hernández MA, et al. Synergistic effects of sulfated polysaccharides from Mexican seaweeds against measles virus. BioMed Res Int. 2016;2016:1-11. doi: 10.1155/2016/8502123
- Pekal A, Pyrzynska K. Availability of Some Elements from Different Types of Teas. Nat Prod J. 2013;3(4):292–5. doi: 10 .2174/221031550304140328113945
- Cuspidi C, Tadic M, Grassi G, Mancia G. Treatment of hypertension: The ESH/ESC guidelines recommendations. Pharmacol Res. 2018;128:315–21. doi: 10.1016/j. phrs.2017.10.003
- 95. Whelton PK, Carey RM, Aronow WS, Casey DE, Collins KJ, Dennison Himmelfarb C, et al. 2017 ACC/AHA/ AAPA/ABC/ACPM/AGS/APhA/ASH/ASPC/NMA/ PCNA guideline for the prevention, detection, evaluation, and management of high blood pressure in adults: a report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. J Am Coll Cardiol. 2018;71(19):e127–248. doi: 10.1161/ HYP.000000000000066.
- 96. Flack JM, Adekola B. Blood pressure and the new ACC/ AHA hypertension guidelines. Trends Cardiovasc Med. 2020;30(3):160-4. doi: 10.1016/j.tcm.2019.05.003
- 97. Gertsch J. Botanical drugs, synergy, and network pharmacology: forth and back to intelligent mixtures. Planta Med. 2011;77(11):1086–98. doi: 10.1055/s-0030-1270904
- 98. Nogales C, Mamdouh ZM, List M, Kiel C, Casas AI, Schmidt HH. Network pharmacology: curing causal mechanisms

#### Saifudin et al

instead of treating symptoms. Trends Pharmacol Sci. 2022;43(2):136–50. doi: 10.1016/j.tips.2021.11.004

- Klein-Junior LC, de Souza MR, Viaene J, Bresolin TM, de Gasper AL, Henriques AT, et al. Quality control of herbal medicines: From traditional techniques to state-of-theart approaches. Planta Med. 2021;87(12/13):964–88. doi: 10.1055/a-1529-8339
- 100. Kim HK, Choi YH, Verpoorte R. NMR-based plant metabolomics: where do we stand, where do we go? Trends Biotechnol. 2011;29(6):267–75. doi: 10.1016/j. tibtech.2011.02.001
- 101. Edison AS, Colonna M, Gouveia GJ, Holderman NR, Judge MT, Shen X, et al. NMR: unique strengths that enhance modern metabolomics research. Anal Chem. 2020;93(1):478–99. doi: 10.1021/acs.analchem.0c04414
- 102. Büscher JM, Czernik D, Ewald JC, Sauer U, Zamboni N. Cross-platform comparison of methods for quantitative metabolomics of primary metabolism. Anal Chem. 2009;81(6):2135–43. doi: 10.1021/ac8022857
- 103. Mukherjee PK, Harwansh RK, Bahadur S, Biswas S, Kuchibhatla LN, Tetali SD, et al. Metabolomics of medicinal plants-a versatile tool for standardization of herbal products and quality evaluation of ayurvedic formulations. Curr Sci. 2016;1624–30. doi: 10.18520/cs/v111/i10/1624-1630
- 104. Mumtaz MW, Hamid AA, Akhtar MT, Anwar F, Rashid U, AL-Zuaidy MH. An overview of recent developments in metabolomics and proteomics-phytotherapic research perspectives. Front Life Sci. 2017;10(1):1–37. doi: 10.1080/21553769.2017.1279573
- 105. Ghosson H, Schwarzenberg A, Jamois F, Yvin JC. Simultaneous untargeted and targeted metabolomics profiling of underivatized primary metabolites in sulfur-deficient barley by ultra-high performance liquid chromatography-quadrupole/time-of-flight mass spectrometry. Plant Methods. 2018;14:1–17. doi: 10.1186/ s13007-018-0329-0
- 106. Caesar LK, Kellogg JJ, Kvalheim OM, Cech NB. Opportunities and limitations for untargeted mass spectrometry metabolomics to identify biologically active constituents in complex natural product mixtures. J Nat Prod. 2019;82(3):469–84. doi: 10.1021/acs. jnatprod.9b00176
- 107. Iglesias Pastrana C, Delgado Bermejo JV, Sgobba MN, Navas González FJ, Guerra L, Pinto DC, et al. Camel (*Camelus* spp.) Urine Bioactivity and Metabolome: A Systematic Review of Knowledge Gaps, Advances, and Directions for Future Research. Int J Mol Sci. 2022;23(23):15024. doi: 10.3390/ijms232315024
- 108. Onuh JO, Aluko RE. Metabolomics as a tool to study the mechanism of action of bioactive protein hydrolysates and peptides: A review of current literature. Trends Food Sci Technol. 2019;91:625–33. doi: 10.1016/j.tifs.2019.08.002
- 109. Chen X, Han W, Wang G, Zhao X. Application prospect of polysaccharides in the development of anti-novel coronavirus drugs and vaccines. Int J Biol Macromol. 2020;164:331–43. doi: 10.1016/j.ijbiomac.2020.07.106
- 110. Xu A, Lai W, Chen P, Awasthi MK, Chen X, Wang Y, et al. A comprehensive review on polysaccharide conjugates derived from tea leaves: Composition, structure, function and application. Trends Food Sci Technol. 2021;114(1):83–

99. doi: 10.1016/j.tifs.2021.05.020

- 111. Zhu C, Li X, Zhang B, Lin Z. Quantitative analysis of multicomponents by single marker-a rational method for the internal quality of Chinese herbal medicine. Integr Med Res. 2017;6(1):1-11. doi: 10.1016/j.imr.2017.01.008
- 112. Kim JH, Lee K, Jerng UM, Choi G, others. Global comparison of stability testing parameters and testing methods for finished herbal products. Evid Based Complement Alternat Med. 2019:1-14. doi: 10.1155/2019/7348929
- 113. Zhu C, Li X, Zhang B, Lin Z. Quantitative analysis of multicomponents by single marker—a rational method for the internal quality of Chinese herbal medicine. Integr Med Res. 2017;6(1):1–11. doi: 10.1016/j.imr.2017.01.008
- 114. Moreira LN, Silva GC, Câmara DV, Pádua RM, Lemos VS, Braga FC, et al. The Cyclitol L-(+)-bornesitol as an active marker for the cardiovascular activity of the brazilian medicinal plant *Hancornia speciosa*. Biol Pharm Bull. 2019;42(12):2076–82. doi: 10.1248/bpb.b19-00601
- 115. Park JH, Whang WK. Bioassay-guided isolation of anti-Alzheimer active components from the aerial parts of *Hedyotis diffusa* and simultaneous analysis for marker compounds. Molecules. 2020;25(24):5867. doi: 10.3390/ molecules25245867
- 116. Länger R, Stöger E, Kubelka W, Helliwell K. Quality standards for herbal drugs and herbal drug preparations– appropriate or improvements necessary? Planta Med. 2018;84(06/07):350–60. doi: 10.1055/s-0043-118534
- 117. Aszyk J, Byliński H, Namieśnik J, Kot-Wasik A. Main strategies, analytical trends and challenges in LC-MS and ambient mass spectrometry-based metabolomics. TrAC Trends Anal Chem. 2018;108:278–95. doi: 10.1016/j. trac.2018.09.010
- 118. Chaleckis R, Meister I, Zhang P, Wheelock CE. Challenges, progress and promises of metabolite annotation for LC-MS-based metabolomics. Curr Opin Biotechnol. 2019;55:44–50. doi: 10.1016/j.copbio.2018.07.010
- 119. Harrieder EM, Kretschmer F, Böcker S, Witting M. Current state-of-the-art of separation methods used in LC-MS based metabolomics and lipidomics. J Chromatogr B. 2022;1188:123069. doi: 10.1016/j.jchromb.2021.123069
- 120. Emwas AH, Roy R, McKay RT, Tenori L, Saccenti E, Gowda GN, et al. NMR spectroscopy for metabolomics research. Metabolites. 2019;9(7):123. doi: 10.3390/metabo9070123
- 121. Giraudeau P. NMR-based metabolomics and fluxomics: developments and future prospects. Analyst. 2020; 145(7):2457–72. doi: 10.1039/D0AN00142B
- Wishart DS. NMR metabolomics: A look ahead. J Magn Reson. 2019;306:155–61. doi: 10.1016/j.jmr.2019.07.013
- 123. Rasdi FLM, Bakar NKA, Mohamad S. A comparative study of selected trace element content in Malay and Chinese traditional herbal medicine (THM) using an inductively coupled plasma-mass spectrometer (ICP-MS). Int J Mol Sci. 2013;14(2):3078–93. doi: 10.3390/ijms14023078
- 124. Sun J, He Y, Yu C, Wang N, Tian L. Elemental Analysis of Xinjiang Rose Hips by Inductively Coupled Plasma-Mass Spectrometry (ICP-MS) and Chemometric Analysis. Anal Lett. 2022;55(2):292–304. doi: 10.1080/00032719.2021.1925904
- 125. United States Pharmacopeia. General Chapter: Assessment of Drug Product Performance—Bioavailability,

Bioequivalence, and Dissolution. USP-NF. Rockville; 2023. p. 1090. doi: 10.31003/USPNF\_M99809\_04\_

- 126. Foster D, Cornella T. Colorimetric Method of Analysis. D. Van Nestrant Company Inc; 1967. p. 551-552.
- 127. Burney CM, Sieburth, JM. Dissolved carbohydrates in seawater. II, A spectrophotometric procedure for total carbohydrate analysis and polysaccharide estimation. Marine Chemistry, 1977;5(1):15-28. doi: 10.1016/0304-4203(77)90012-3
- 128. Wu L, Gao Y, Ren W chen, Su Y, Li J, Du Y qi, et al. Rapid determination and origin identification of total polysaccharides contents in Schisandra chinensis by nearinfrared spectroscopy. Spectrochim Acta A Mol Biomol Spectrosc. 2022;264:120327. doi: 10.1016/j.saa.2021.120327
- 129. Sukhija PS, Palmquist D. Rapid method for determination of total fatty acid content and composition of feedstuffs and feces. J Agric Food Chem. 1988;36(6):1202–6. doi: 10.1021/ jf00084a019
- 130. Eg B. A rapid method of total lipid extraction and purification. Can J Biochem Physiol. 1959;39:911–7. doi: 10.1139/o59-099
- Cavonius LR, Carlsson NG, Undeland I. Quantification of total fatty acids in microalgae: comparison of extraction and transesterification methods. Anal Bioanal Chem. 2014;406:7313–22. doi: 10.1007/s00216-014-8155-3
- Stuart NW. Adaptation of the micro-Kjeldahl method for the determination of nitrogen in plant tissues. Plant Physiol. 1936;11(1):173. doi: 10.1104/pp.11.1.173
- 133. Bowman D, Paul J, Carlson R. A method to exclude nitrate from Kjeldahl digestion of plant tissues. Commun Soil Sci Plant Anal. 1988;19(2):205–13. doi: 10.1080/00103628809367932
- Méchin V, Damerval C, Zivy M. Total protein extraction with TCA-acetone. Plant Proteomics Methods Protoc. 2007;1–8. doi: 10.1385/1-59745-227-0:1
- 135. Erdemir US. Contribution of tea (*Camellia sinensis* L.) to recommended daily intake of Mg, Mn, and Fe: An in vitro bioaccessibility assessment. J Food Compos Anal. 2018;69:71–7. doi: 10.1016/j.jfca.2018.02.006
- 136. Junior JBP, Dantas KG. Evaluation of inorganic elements in cat's claw teas using ICP OES and GF AAS. Food Chem. 2016;196:331–7. doi: 10.1016/j.foodchem.2015.09.057
- 137. Rao Y, Xiang B. Determination of total ash and acidinsoluble ash of Chinese herbal medicine Prunellae Spica by near infrared spectroscopy. Yakugaku Zasshi. 2009;129(7):881–6. doi: 10.1248/yakushi.129.881
- Matić P, Sabljić M, Jakobek L. Validation of spectrophotometric methods for the determination of total polyphenol and total flavonoid content. J AOAC Int. 2017;100(6):1795–803. doi: 10.5740/jaoacint.17-0066
- 139. Shraim AM, Ahmed TA, Rahman MM, Hijji YM. Determination of total flavonoid content by aluminum chloride assay: A critical evaluation. Lwt. 2021;150:111932. doi: 10.1016/j.lwt.2021.111932
- 140. Poláček R, Májek P, Hroboňová K, Sádecká J. Fluorescence

spectroscopy as a tool for determination of coumarins by multivariate calibration. J Fluoresc. 2015;25:297–303. doi: 10.1007/s10895-015-1508-2

- 141. Baccou J, Lambert F, Sauvaire Y. Spectrophotometric method for the determination of total steroidal sapogenin. Analyst. 1977;102(1215):458–65. doi: 10.1039/an9770200458
- 142. Rai M, Ramachandran K, Gupta V. Spectrophotometric method for the determination of total tobacco alkaloids and nicotine. Analyst. 1994;119(8):1883–5. doi: 10.1039/ an9941901883
- 143. Hu T, He XW, Jiang JG, Xu XL. Efficacy evaluation of a Chinese bitter tea (*Ilex latifolia* Thunb.) via analyses of its main components. Food Funct. 2014;5(5):876–81. doi: 10.1039/c3fo60603a
- 144. Lee CH, Lee TH, Ong PY, Wong SL, Hamdan N, Elgharbawy AA, et al. Integrated ultrasound-mechanical stirrer technique for extraction of total alkaloid content from *Annona muricata*. Process Biochem. 2021;109:104– 16. doi: 10.1016/j.procbio.2021.07.006
- 145. Font R, del Río-Celestino M, Cartea E, de Haro-Bailón A. Quantification of glucosinolates in leaves of leaf rape (*Brassica napus* ssp. pabularia) by near-infrared spectroscopy. Phytochemistry. 2005;66(2):175–85. doi: 10.1016/j.phytochem.2004.11.011
- 146. Cartea ME, Velasco P, Obregón S, Padilla G, de Haro A. Seasonal variation in glucosinolate content in *Brassica oleracea* crops grown in northwestern Spain. Phytochemistry. 2008;69(2):403–10. doi: 10.1016/j. phytochem.2007.08.014
- 147. Cheng D, Vrieling K, Klinkhamer PG. The effect of hybridization on secondary metabolites and herbivore resistance: implications for the evolution of chemical diversity in plants. Phytochem Rev. 2011;10:107–17. doi: 10.1007/s11101-010-9194-9
- 148. Wöll S, Kim SH, Greten HJ, Efferth T. Animal plant warfare and secondary metabolite evolution. Nat Prod Bioprospecting. 2013;3:1–7. doi: 10.1007/s13659-013-0004-0
- 149. Deduke C, Timsina B, Piercey-Normore MD. Effect of environmental change on secondary metabolite production in lichen-forming fungi. In: International Perspectives on Global Environmental Change. IntechOpen; 2012. p. 197-230. doi: 10.5772/26954
- 150. Li Y, Kong D, Fu Y, Sussman MR, Wu H. The effect of developmental and environmental factors on secondary metabolites in medicinal plants. Plant Physiol Biochem. 2020;148:80–9. doi: 10.1016/j.plaphy.2020.01.006
- 151. Chen D, Mubeen B, Hasnain A, Rizwan M, Adrees M, Naqvi SAH, et al. Role of promising secondary metabolites to confer resistance against environmental stresses in crop plants: Current scenario and future perspectives. Front Plant Sci. 2022;13:881032. doi: 10.3389/fpls.2022.881032
- 152. Kessler A, Kalske A. Plant secondary metabolite diversity and species interactions. Annu Rev Ecol Evol Syst. 2018;49:115– 38.doi:10.1146/annurev-ecolsys-110617-062406